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**UNITED STATES DISTRICT COURT
DISTRICT OF NEW JERSEY**

NOVO NORDISK INC. and
NOVO NORDISK A/S,

Plaintiffs,

v.

RIO BIOPHARMACEUTICALS, INC.,

Defendant.

C.A. No. _____

(Filed Electronically)

COMPLAINT FOR PATENT INFRINGEMENT

Plaintiffs Novo Nordisk Inc. and Novo Nordisk A/S (collectively, “Novo Nordisk”), by their undersigned attorneys, bring this action against Defendant Rio Biopharmaceuticals, Inc. (“Rio”), and hereby allege as follows:

NATURE OF THE ACTION

1. This is an action for patent infringement under the patent laws of the United States, Title 35 of the United States Code, arising from Rio's submission of an Abbreviated New Drug Application ("ANDA") to the United States Food and Drug Administration ("FDA"), by which Rio seeks approval to market a generic version of Novo Nordisk's pharmaceutical product Victoza[®] prior to the expiration of United States Patent Nos. 8,114,833 (the "'833 patent") and 9,265,893 (the "'893 patent"), which cover, *inter alia*, Victoza[®] and/or its use.

THE PARTIES

2. Plaintiff Novo Nordisk Inc. ("NNI") is a corporation organized and existing under the laws of the State of Delaware and has its principal place of business at 800 Scudders Mill Road, Plainsboro, New Jersey 08536.

3. Plaintiff Novo Nordisk A/S ("NNAS") is an entity organized and existing under the laws of the Kingdom of Denmark and has its principal place of business at Novo Allé, 2880 Bagsværd, Denmark. NNI is an indirect, wholly owned subsidiary of NNAS.

4. On information and belief, Rio is a corporation organized and existing under the laws of the State of Delaware, having its principal place of business at 116 Village Blvd, Suite 200, Princeton, NJ 08540. On information and belief, Rio is in the business of making and selling generic pharmaceutical products, for distribution in the State of New Jersey and throughout the United States.

JURISDICTION AND VENUE

5. This action for patent infringement arises under 35 U.S.C. § 1 *et seq.* generally and 35 U.S.C. § 271 specifically.

6. This Court has subject matter jurisdiction over this dispute pursuant to 28 U.S.C. §§ 1331 and 1338(a).

7. Venue is proper in this Judicial District pursuant to 28 U.S.C. §§ 1391 and 1400(b).

8. This Court has personal jurisdiction over Rio because, upon information and belief, it has a physical presence in New Jersey; it conducts business in New Jersey; it derives revenue from conducting business in New Jersey; and it has engaged in systematic and continuous contacts with the State of New Jersey, either directly or through its affiliates and/or agents, including by marketing and/or selling pharmaceutical products in New Jersey, including in this Judicial District.

9. On information and belief, Rio intends to sell, offer to sell, use, and/or engage in the commercial manufacture of a generic version of liraglutide injection solution, 18 mg/3 ml (6 mg/ml) (“Rio’s Product”), directly or indirectly, throughout the United States and in this Judicial District. Rio’s filing of Rio’s ANDA No. 218241 (“Rio’s ANDA”) confirms this intention and further subjects Rio to the specific personal jurisdiction of this Court.

THE PATENTS-IN-SUIT

10. On February 14, 2012, the United States Patent and Trademark Office issued the ’833 patent, entitled, “Propylene Glycol-Containing Peptide Formulations Which Are Optimal for Production and For Use in Injection Devices,” a copy of which is attached to this Complaint as Exhibit A. NNAS is the owner of all right, title, and interest in the ’833 patent.

11. On February 23, 2016, the United States Patent and Trademark Office issued the ’893 patent, entitled, “Injection Button,” a copy of which is attached to this Complaint as Exhibit B. NNAS is the owner of all right, title, and interest in the ’893 patent.

VICTOZA®

12. NNI holds approved New Drug Application No. 022341 (the “Victoza® NDA”) for Liraglutide Recombinant Solution Injection, 18 mg/3 ml (6 mg/ml), which NNI sells under the trade name Victoza®.

13. The claims of the patents-in-suit cover, *inter alia*, Victoza® and/or its use.

14. Pursuant to 21 U.S.C. § 355(b)(1), and attendant FDA regulations, the ’833 and ’893 patents are listed in the FDA publication, “Approved Drug Products with Therapeutic Equivalence Evaluations” (the “Orange Book”), with respect to Victoza®.

RIO’S ANDA

15. On information and belief, Rio submitted Rio’s ANDA to the FDA, pursuant to 21 U.S.C. § 355(j), seeking approval to market Rio’s Product, which is a generic version of liraglutide injection solution, 18 mg/3 ml (6 mg/ml).

16. On information and belief, Rio’s ANDA refers to and relies upon the Victoza® NDA and contains data that, according to Rio, demonstrate the bioequivalence of Rio’s Product and Victoza®.

17. By letter to NNI and NNAS dated December 8, 2023 (the “Notice Letter”), Rio stated that Rio’s ANDA contained a certification pursuant to 21 U.S.C. § 355(j)(2)(A)(vii)(IV) that the ’833 and ’893 patents are invalid, unenforceable, and/or will not be infringed by the commercial manufacture, use, or sale of Rio’s Product (the “Paragraph IV Certification”). Rio attached a memorandum to the Notice Letter in which it purported to allege factual and legal bases for its Paragraph IV Certification. NNI and NNAS file this suit within 45 days of receipt of the Notice Letter.

COUNT I: INFRINGEMENT OF U.S. PATENT NO. 8,114,833

18. Novo Nordisk re-alleges and incorporates by reference the allegations of Paragraphs 1–17 of this Complaint.

19. Rio has infringed the '833 patent, pursuant to 35 U.S.C. § 271(e)(2)(A), by submitting Rio's ANDA, by which Rio seeks approval from the FDA to manufacture, use, offer to sell, and sell Rio's Product prior to the expiration of the '833 patent.

20. Claims 1–15 of the '833 patent are directed to GLP-1 formulations. Claims 16–31 are directed to methods for preparing such formulations or methods of reducing deposits or reducing clogging by replacing the isotonicity agent in a formulation with propylene glycol. Rio's manufacture, use, offer to sell, or sale of Rio's Product within the United States, or importing of Rio's Product into the United States, during the term of the '833 patent would infringe claims 1–31 of the '833 patent.

21. Novo Nordisk will be harmed substantially and irreparably if Rio is not enjoined from infringing the '833 patent and/or if the FDA is not enjoined from approving Rio's ANDA before the '833 patent expires.

22. Novo Nordisk has no adequate remedy at law.

23. Rio was aware of the '833 patent when it submitted its ANDA. Novo Nordisk is entitled to a finding that this case is exceptional and to an award of attorneys' fees under 35 U.S.C. § 285.

COUNT II: INFRINGEMENT OF U.S. PATENT NO. 9,265,893

24. Novo Nordisk re-alleges and incorporates by reference the allegations of Paragraphs 1–23 of this Complaint.

25. Rio has infringed the '893 patent, pursuant to 35 U.S.C. § 271(e)(2)(A), by submitting Rio's ANDA, by which Rio seeks approval from the FDA to manufacture, use, offer to sell, and sell Rio's Product prior to the expiration of the '893 patent.

26. Claims 1–6 of the '893 patent are directed to a push button connection for an injection device. Rio's manufacture, use, offer for sale, or sale of Rio's Product within the United States, or importing of Rio's Product into the United States, during the term of the '893 patent would infringe claims 1–6 of the '893 patent.

27. Novo Nordisk will be harmed substantially and irreparably if Rio is not enjoined from infringing the '893 patent and/or if the FDA is not enjoined from approving Rio's ANDA before the '893 patent expires.

28. Novo Nordisk has no adequate remedy at law.

29. Rio was aware of the '893 patent when it submitted its ANDA. Novo Nordisk is entitled to a finding that this case is exceptional and to an award of attorneys' fees under 35 U.S.C. § 285.

PRAYER FOR RELIEF

WHEREFORE, Novo Nordisk prays for a judgment in its favor and against Rio and respectfully requests the following relief:

- A. A Judgment that Rio has infringed the '833 patent;
- B. A Judgment that Rio has infringed the '893 patent;
- C. A Judgment ordering that, pursuant to 35 U.S.C. § 271(e)(4)(A), the effective date of any approval of Rio's ANDA, under § 505(j) of the Federal Food, Drug, and Cosmetic Act (21 U.S.C. § 355(j)), shall not be earlier than the expiration of the '833 and '893 patents, including

any extensions, adjustments, and exclusivities for those patents to which Novo Nordisk is or becomes entitled;

D. A Judgment, pursuant to 35 U.S.C. § 271(e)(4)(B), preliminarily and permanently enjoining Rio, its officers, agents, servants, and employees, and those persons in active concert or participation with any of them, from manufacturing, using, offering to sell, or selling Rio's Product within the United States, or importing Rio's Product into the United States, prior to the expiration of the '833 and '893 patents, including any extensions, adjustments, and exclusivities for those patents to which Novo Nordisk is or becomes entitled;

E. If Rio commercially manufactures, uses, offers to sell, or sells Rio's Product within the United States, or imports Rio's Product into the United States, prior to the expiration of the '833 patent and '893 patent, including any extensions, adjustments, and exclusivities, for those patents to which Novo Nordisk is or becomes entitled, a Judgment awarding Novo Nordisk monetary relief, including, but not limited to, lost profits, together with interest;

F. Attorneys' fees in this action as an exceptional case pursuant to 35 U.S.C. § 285;

G. Costs and expenses in this action; and

H. Such other relief as the Court deems just and proper.

Dated: January 19, 2024

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CERTIFICATION PURSUANT TO LOCAL CIVIL RULES 11.2 & 40.1

I hereby certify that the matters *Novo Nordisk Inc., et al. v. Lupin Ltd.*, Civil Action No. 23-4027 (RMB)(SAK) (D.N.J.); *Novo Nordisk Inc., et al. v. Lupin Ltd.*, Civil Action No. 23-4031 (RMB)(SAK) (D.N.J.); *Novo Nordisk Inc., et al. v. ScinoPharm Taiwan Ltd.*, Civil Action No. 23-20935 (RMB)(SAK) (D.N.J.); and *Novo Nordisk Inc. et al. v. Dr. Reddy's Laboratories, Ltd. et al.*, C.A. No. 23-22112 (RMB)(SAK) (D.N.J.), are related to the matter in controversy because they involve the same plaintiffs, some of the same patents, and Rio is seeking FDA approval to market generic versions of the same or similar pharmaceutical products.

I hereby certify that the matter in controversy is also related to the following matters because they involve the same plaintiffs and some of the same patents: *Novo Nordisk Inc., et al. v. Biocon Pharma Ltd., et al.*, C.A. No. 22-937 (CFC) (D. Del.); *Novo Nordisk Inc., et al. v. Orbicular Pharm. Technologies Pvt. Ltd.*, C.A. Nos. 22-856 (CFC) and 23-179 (CFC) (D. Del.).

I hereby certify that, to the best of my knowledge, the matter in controversy is not the subject of any other action pending in any court or of any arbitration or administrative proceeding.

Dated: January 19, 2024

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EXHIBIT A



US008114833B2

(12) **United States Patent**
Pedersen et al.

(10) **Patent No.:** **US 8,114,833 B2**
(45) **Date of Patent:** ***Feb. 14, 2012**

(54) **PROPYLENE GLYCOL-CONTAINING
PEPTIDE FORMULATIONS WHICH ARE
OPTIMAL FOR PRODUCTION AND FOR USE
IN INJECTION DEVICES**

(75) Inventors: **Tina Bjeldskov Pedersen**, Smørum
(DK); **Claude Bonde**, Lyngby (DK);
Dorthe Kot Englund, Holte (DK)

(73) Assignee: **Novo Nordisk A/S**, Bagsvaerd (DK)

(*) Notice: Subject to any disclaimer, the term of this
patent is extended or adjusted under 35
U.S.C. 154(b) by 663 days.

This patent is subject to a terminal dis-
claimer.

(21) Appl. No.: **11/435,977**

(22) Filed: **May 17, 2006**

(65) **Prior Publication Data**

US 2007/0010424 A1 Jan. 11, 2007

Related U.S. Application Data

(63) Continuation of application No.
PCT/DK2004/000792, filed on Nov. 18, 2004.

(60) Provisional application No. 60/524,653, filed on Nov.
24, 2003.

(30) **Foreign Application Priority Data**

Nov. 20, 2003 (DK) 2003 01719

(51) **Int. Cl.**
A61K 38/26 (2006.01)

(52) **U.S. Cl.** **514/2**; 530/308

(58) **Field of Classification Search** None
See application file for complete search history.

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Primary Examiner — Christina Bradley

(74) *Attorney, Agent, or Firm* — Michael J. Brignati

(57) **ABSTRACT**

The present invention relates to pharmaceutical formulations comprising a peptide and propylene glycol, to methods of preparing such formulations, and to uses of such formulations in the treatment of diseases and conditions for which use of the peptide contained in such formulations is indicated. The present invention further relates to methods for reducing the clogging of injection devices by a peptide formulation and for reducing deposits on production equipment during production of a peptide formulation.

31 Claims, 7 Drawing Sheets

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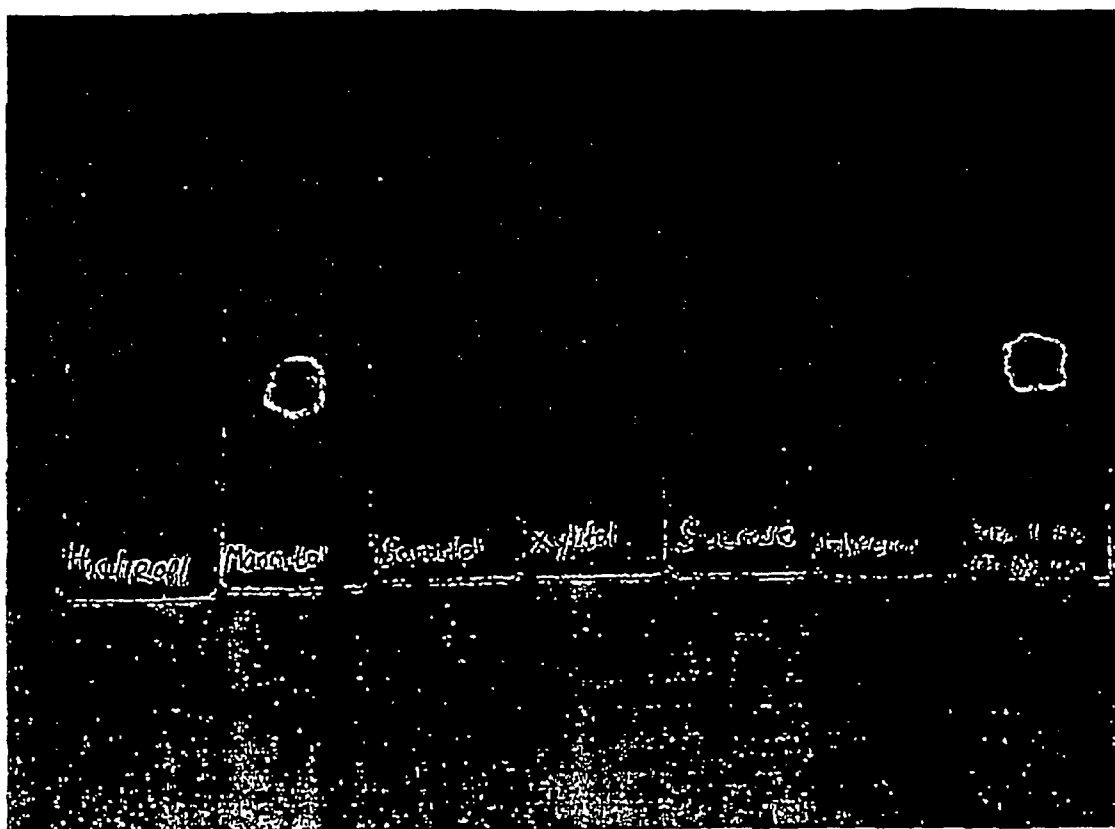
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FIGURE 1



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FIGURE 2



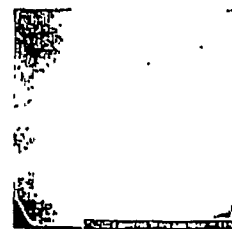
Mannitol



Argi-



Inosi-



Glyce-

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FIGURE 3



Myo-inositol



Maltose



Glycerol

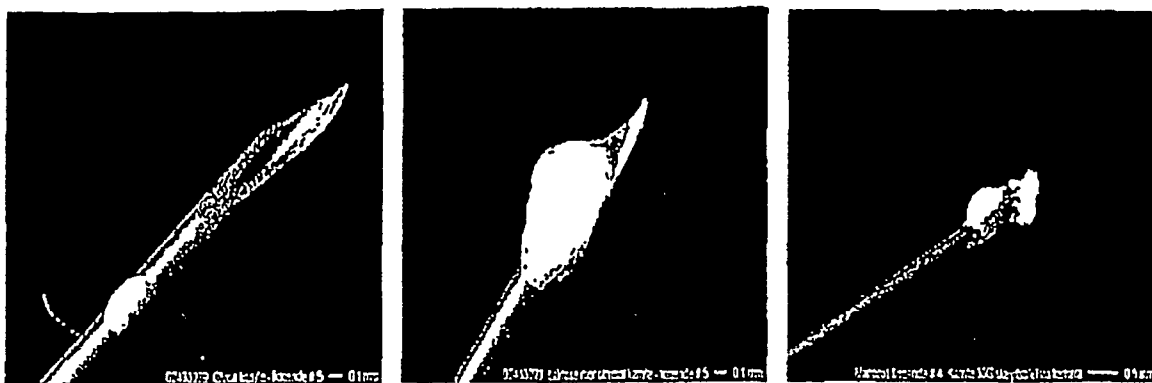
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FIGURE 4



Glycine

Lactose

Mannitol

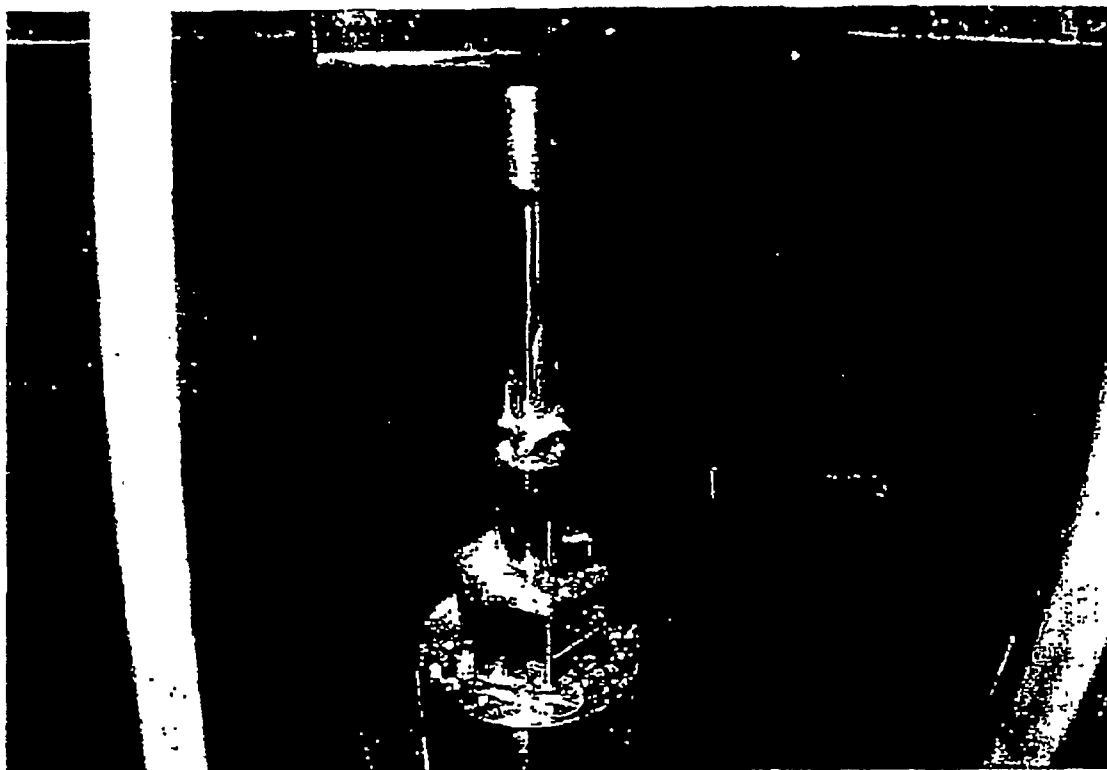
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FIGURE 5



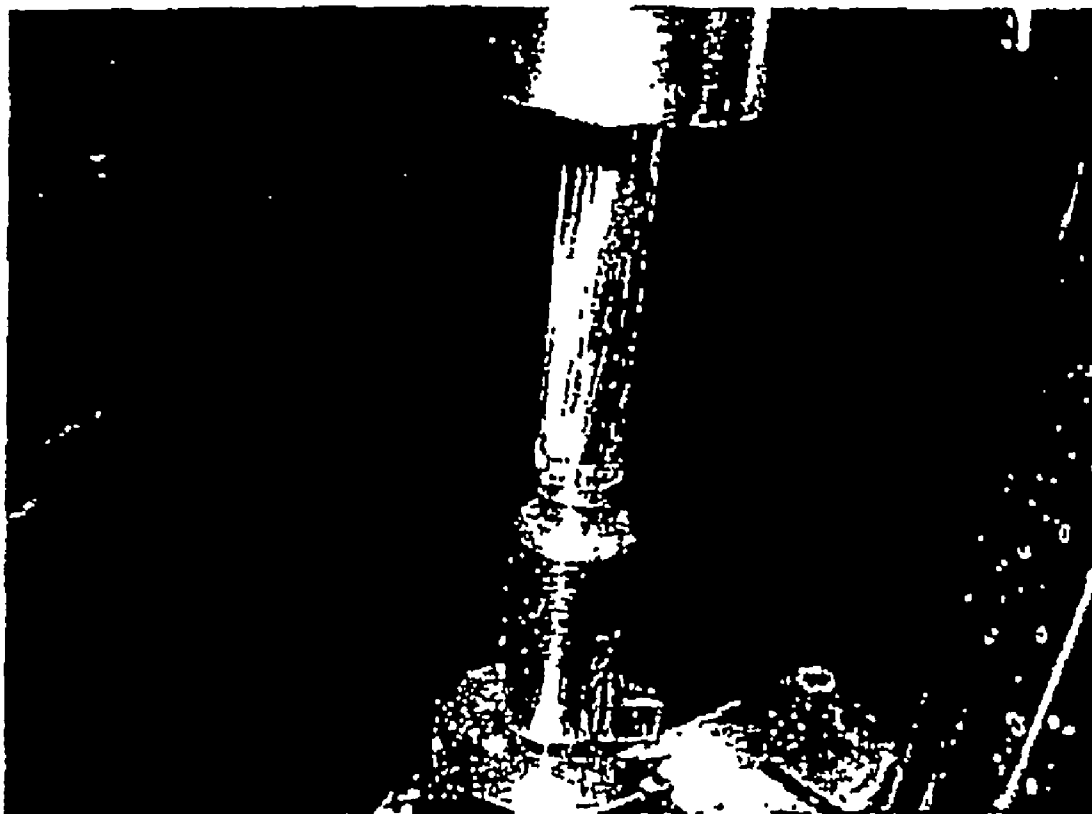
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FIGURE 6



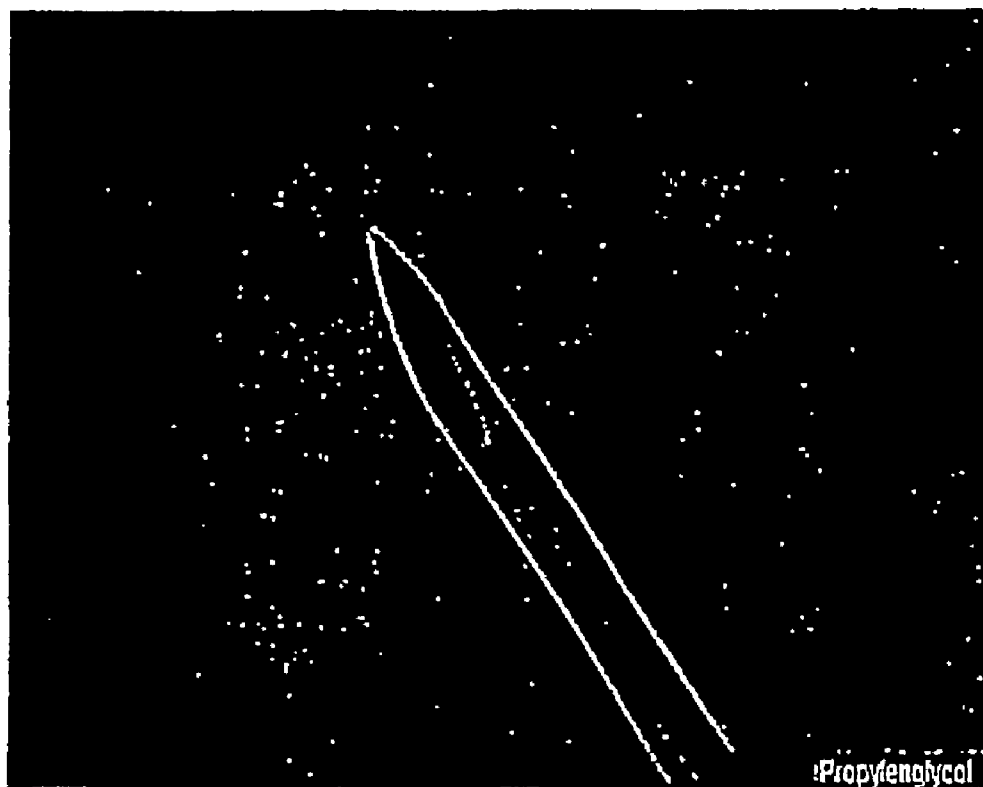
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FIGURE 7



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**PROPYLENE GLYCOL-CONTAINING
PEPTIDE FORMULATIONS WHICH ARE
OPTIMAL FOR PRODUCTION AND FOR USE
IN INJECTION DEVICES**

CROSS REFERENCE TO RELATED
APPLICATIONS

This Application is a continuation of International Application serial no. PCT/DK2004/000792 filed Nov. 18, 2004 and claims priority from U.S. application Ser. No. 60/524,653 filed Nov. 24, 2003 and from Danish Application serial no. PA 2003 01719 filed Nov. 20, 2003.

FIELD OF THE INVENTION

The present invention relates to pharmaceutical formulations comprising a peptide and propylene glycol, to methods of preparing such formulations, and to uses of such formulations in the treatment of diseases and conditions for which use of the peptide contained in such formulations is indicated. The present invention further relates to methods for reducing the clogging of injection devices by a peptide formulation and for reducing deposits on production equipment during production of a peptide formulation.

BACKGROUND OF THE INVENTION

The inclusion of isotonicity agents in peptide-containing pharmaceutical formulations is widely known and one of the more common isotonic agents used in such formulations is mannitol. However, the present inventors have observed that mannitol causes problems during the production of peptide formulations as it crystallizes resulting in deposits in the production equipment and in the final product. Such deposits increase the need to clean the filling equipment during production of the formulation and this results in reduced production capability. In addition, such deposits may also result in reduced yield of the final product since vials/cartridges containing the peptide formulation may need to be discarded if particles are present. Finally, the present inventors have observed that in peptide formulations to be administered by injection, the presence of mannitol results in clogging of injection devices.

Accordingly, it is desirable to identify an alternative isotonic agent to mannitol for inclusion in peptide-containing formulations and in particular, for inclusion in peptide formulations which are administered by injection.

SUMMARY OF THE INVENTION

The present inventors have discovered that peptide formulations containing propylene glycol at certain concentrations exhibit reduced deposits in production equipment and in the final product and also exhibit reduced clogging of injection devices. The present compositions may be formulated with any peptide and are also physically and chemically stable thus rendering them shelf-stable and suitable for invasive (e.g. injection, subcutaneous injection, intramuscular, intravenous or infusion) as well as non-invasive (e.g. nasal, oral, pulmonary, transdermal or transmucosal e.g. buccal) means of administration.

The present invention therefore relates to a pharmaceutical formulation comprising a peptide and propylene glycol, where the propylene glycol is present in a concentration of 1-100 mg/ml and the pH of the formulation is from 7-10. In a

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preferred embodiment, the pharmaceutical formulations of the invention further contain a buffer and a preservative.

The present invention also relates to methods for producing the pharmaceutical formulations of the invention.

5 In one embodiment, the method for preparing a peptide formulation comprises:

- a) preparing a first solution by dissolving preservative, propylene glycol and buffer in water;
- 10 b) preparing a second solution by dissolving the peptide in water;
- c) mixing the first and second solutions; and
- d) adjusting the pH of the mixture in c) to the desired pH.

In another embodiment, the method for preparing a peptide formulation comprises:

- 15 a) preparing a first solution by dissolving preservative and buffer in water;
- b) adding propylene glycol to the first solution;
- c) mixing the first solution with a second solution containing peptide dissolved in water; and
- 20 d) adjusting the pH of the mixture in c) to the desired pH.

In yet another embodiment, the method for preparing a peptide formulation comprises:

- 25 a) preparing a solution by dissolving preservative, buffer and propylene glycol in water;
- b) adding the peptide to the solution of step a); and
- c) adjusting the pH of the solution of step b) to the desired pH.

The present invention further relates to methods of treatment using the pharmaceutical formulations of the invention where the compositions are administered in an amount effective to combat the disease, condition, or disorder for which administration of the peptide contained in the formulation is indicated.

30 In addition the present invention also relates to a method for reducing deposits on production equipment during production of a peptide formulation, where the method comprises replacing the isotonicity agent previously utilized in said formulation with propylene glycol at a concentration of between 1-100 mg/ml.

In one embodiment, the reduction in deposits on the production equipment during production by the propylene glycol-containing formulation relative to that observed for the formulation containing the previously utilized isotonicity agent is measured by a simulated filling experiment.

45 The present invention also relates to a method for reducing deposits in the final product during production of a peptide formulation, where the method comprises replacing the isotonicity agent previously utilized in said formulation with propylene glycol at a concentration of between 1-100 mg/ml.

In one embodiment, the reduction in deposits in the final product is measured by a reduction in the number of vials and/or cartridges of the propylene glycol-containing formulation that must be discarded due to deposits relative to number of vials and/or cartridges of the formulation containing the previously utilized isotonicity agent that must be discarded due to deposits.

50 The present invention further relates to a method for reducing the clogging of injection devices by a peptide formulation, where the method comprises replacing the isotonicity agent previously utilized in said formulation with propylene glycol at a concentration of between 1-100 mg/ml.

In one embodiment, the reduction in clogging of the injection device by the propylene glycol-containing formulation relative to that observed for the formulation containing the previously utilized isotonicity agent is measured in a simulated in use study.

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BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 shows a photograph of dried droplets on microscope slides of from left to right, placebo (no peptide) formulations containing no isotonic agent (e only water, preservative and buffer), mannitol, sorbitol, xylitol, sucrose or glycerol as the isotonic agent with the far right slide containing mannitol with peptide Arg³⁴, Lys²⁶(N^ε-(γ-Glu(N^α-hexadecanoyl)))-GLP-1(7-37).

FIG. 2 shows light microscopy pictures of from left to right, some of the dried droplets of placebo formulations containing mannitol, arginin, inositol or glycerol as the isotonic agent.

FIG. 3 shows light microscopy pictures of clogged needles dosed with placebo formulations containing myoinositol, maltose or glycerol as the isotonic agent.

FIG. 4 shows light microscopy pictures of deposits on needles dosed with placebo formulations containing glycine, lactose or mannitol as the isotonic agent.

FIG. 5 shows filling equipment after 24 hours simulated filling with Arg³⁴, Lys²⁶(N^ε-(γ-Glu(N^α-hexadecanoyl)))-GLP-1(7-37) medium containing myo-inositol.

FIG. 6 shows deposits on filling equipment after 24 hours simulated filling with a mannitol-containing placebo formulation.

FIG. 7 shows deposits on needles dosed with mannitol (top panel) and propylene glycol (bottom panel)-containing Arg³⁴, Lys²⁶(N^ε-(γ-Glu(N^α-hexadecanoyl)))-GLP-1(7-37) formulations.

DESCRIPTION OF THE INVENTION

The present invention relates to a pharmaceutical formulation comprising a peptide or a mixture of peptides and propylene glycol where the final concentration of propylene glycol in the formulation is 1-100 mg/ml and the pH of the formulation is in the range of from 7-10.

The pharmaceutical formulations of the invention are found to be optimal for production because they exhibit reduced deposits in production equipment relative to formulations containing other isotonicity agents as measured by the simulated filling studies described in the Examples. In addition, the pharmaceutical formulations of the invention are found to be optimal for use in injection devices because they exhibit reduced clogging of the injection devices relative to formulations containing other isotonicity agents as measured by the simulated in use studies described in the Examples.

The formulations of the present invention may be formulated with any peptide where examples of such peptides include, but are not limited to, glucagon, human growth hormone (hGH), insulin, aprotinin, FactorVII, tissue plasminogen activator (TPA), FactorVIIa, FFR-FactorVIIa, heparinase, ACTH, Heparin Binding Protein, corticotropin-releasing factor, angio-tensin, calcitonin, glucagon-like peptide-1, glucagon-like peptide-2, insulin-like growth factor-1, insulin-like growth factor-2, fibroblast growth factors, gastric inhibitory peptide, growth hormone-releasing factor, pituitary adenylate cyclase activating peptide, secretin, enterogastrin, somatostatin, somatomedin, parathyroid hormone, thrombopoietin, erythropoietin, hypothalamic releasing factors, prolactin, thyroid stimulating hormones, endorphins, enkephalins, vasopressin, oxytocin, opioids, DPP IV, interleukins, immunoglobulins, complement inhibitors, serine protease inhibitors, cytokines, cytokine receptors, PDGF, tumor necrosis factors, tumor necrosis factors receptors, growth factors and analogues as well as derivatives

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thereof where each of these peptides constitutes an alternative embodiment of the present invention.

In the present application, the designation “an analogue” is used to designate a peptide wherein one or more amino acid residues of the parent peptide have been substituted by another amino acid residue and/or wherein one or more amino acid residues of the parent peptide have been deleted and/or wherein one or more amino acid residues have been added to the parent peptide. Such addition can take place either at the N-terminal end or at the C-terminal end of the parent peptide or both. Typically “an analogue” is a peptide wherein 6 or less amino acids have been substituted and/or added and/or deleted from the parent peptide, more preferably a peptide wherein 3 or less amino acids have been substituted and/or added and/or deleted from the parent peptide, and most preferably, a peptide wherein one amino acid has been substituted and/or added and/or deleted from the parent peptide.

In the present application, “a derivative” is used to designate a peptide or analogue thereof which is chemically modified by introducing an organic substituent e.g. ester, alkyl or lipophilic functionalities, on one or more amino acid residues of the peptide or analogue thereof.

In one embodiment, the peptide to be included in the formulation of the invention is a GLP-1 agonist where “a GLP-1 agonist” is understood to refer to any peptide which fully or partially activates the human GLP-1 receptor. In a preferred embodiment, the “GLP-1 agonist” is any peptide that binds to a GLP-1 receptor, preferably with an affinity constant (K_D) or a potency (EC₅₀) of below 1 μM, e.g. below 100 nM as measured by methods known in the art (see e.g. WO 98/08871) and exhibits insulinotropic activity, where insulinotropic activity may be measured in vivo or in vitro assays known to those of ordinary skill in the art. For example, the GLP-1 agonist may be administered to an animal and the insulin concentration measured over time.

Methods for identifying GLP-1 agonists are described in WO 93/19175 (Novo Nordisk A/S) and examples of suitable GLP-1 analogues and derivatives which can be used according to the present invention includes those referred to in WO 99/43705 (Novo Nordisk A/S), WO 99/43706 (Novo Nordisk A/S), WO 99/43707 (Novo Nordisk A/S), WO 98/08871 (analogues with lipophilic substituent) and in WO 02/46227 (analogues fused to serum albumin or to Fc portion of an Ig). (Novo Nordisk A/S), WO 99/43708 (Novo Nordisk A/S), WO 99/43341 (Novo Nordisk A/S), WO 87/06941 (The General Hospital Corporation), WO 90/11296 (The General Hospital Corporation), WO 91/11457 (Buckley et al.), WO 98/43658 (Eli Lilly & Co.), EP 0708179-A2 (Eli Lilly & Co.), EP 0699686-A2 (Eli Lilly & Co.), WO 01/98331 (Eli Lilly & Co.).

In one embodiment, the GLP-1 agonist is selected from the group consisting of GLP-1(7-36)-amide, GLP-1(7-37), a GLP-1(7-36)-amide analogue, a GLP-1(7-37) analogue, or a derivative of any of these.

In one embodiment, the GLP-1 agonist is a derivative of GLP-1(7-36)-amide, GLP-1(7-37), a GLP-1(7-36)-amide analogue or a GLP-1(7-37) analogue, which comprises a lipophilic substituent.

In this embodiment of the invention, the GLP-1 derivative preferably has three lipophilic substituents, more preferably two lipophilic substituents, and most preferably one lipophilic substituent attached to the parent peptide (ie GLP-1(7-36)-amide, GLP-1(7-37), a GLP-1(7-36)-amide analogue or a GLP-1(7-37) analogue), where each lipophilic substituent (s) preferably has 4-40 carbon atoms, more preferably 8-30

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carbon atoms, even more preferably 8-25 carbon atoms, even more preferably 12-25 carbon atoms, and most preferably 14-18 carbon atoms.

In one embodiment, the lipophilic substituent comprises a partially or completely hydrogenated cyclopentanophenathrene skeleton.

In another embodiment, the lipophilic substituent is a straight-chain or branched alkyl group.

In yet another embodiment, the lipophilic substituent is an acyl group of a straight-chain or branched fatty acid. Preferably, the lipophilic substituent is an acyl group having the formula $\text{CH}_3(\text{CH}_2)_n\text{CO}-$, wherein n is an integer from 4 to 38, preferably an integer from 12 to 38, and most preferably is $\text{CH}_3(\text{CH}_2)_{12}\text{CO}-$, $\text{CH}_3(\text{CH}_2)_{14}\text{CO}-$, $\text{CH}_3(\text{CH}_2)_{16}\text{CO}-$, $\text{CH}_3(\text{CH}_2)_{18}\text{CO}-$, $\text{CH}_3(\text{CH}_2)_{20}\text{CO}-$ and $\text{CH}_3(\text{CH}_2)_{22}\text{CO}-$. In a more preferred embodiment, the lipophilic substituent is tetradecanoyl. In a most preferred embodiment, the lipophilic substituent is hexadecanoyl.

In a further embodiment of the present invention, the lipophilic substituent has a group which is negatively charged such as a carboxylic acid group. For example, the lipophilic substituent may be an acyl group of a straight-chain or branched alkane α,ω -dicarboxylic acid of the formula $\text{HOOC}(\text{CH}_2)_m\text{CO}-$, wherein m is an integer from 4 to 38, preferably an integer from 12 to 38, and most preferably is $\text{HOOC}(\text{CH}_2)_{14}\text{CO}-$, $\text{HOOC}(\text{CH}_2)_{16}\text{CO}-$, $\text{HOOC}(\text{CH}_2)_{18}\text{CO}-$, $\text{HOOC}(\text{CH}_2)_{20}\text{CO}-$ or $\text{HOOC}(\text{CH}_2)_{22}\text{CO}-$.

In the GLP-1 derivatives of the invention, the lipophilic substituent(s) contain a functional group which can be attached to one of the following functional groups of an amino acid of the parent GLP-1 peptide:

- (a) the amino group attached to the alpha-carbon of the N-terminal amino acid,
- (b) the carboxy group attached to the alpha-carbon of the C-terminal amino acid,
- (c) the epsilon-amino group of any Lys residue,
- (d) the carboxy group of the R group of any Asp and Glu residue,
- (e) the hydroxy group of the R group of any Tyr, Ser and Thr residue,
- (f) the amino group of the R group of any Trp, Asn, Gln, Arg, and His residue, or
- (g) the thiol group of the R group of any Cys residue.

In one embodiment, a lipophilic substituent is attached to the carboxy group of the R group of any Asp and Glu residue.

In another embodiment, a lipophilic substituent is attached to the carboxy group attached to the alpha-carbon of the C-terminal amino acid.

In a most preferred embodiment, a lipophilic substituent is attached to the epsilon-amino group of any Lys residue.

In a preferred embodiment of the invention, the lipophilic substituent is attached to the parent GLP-1 peptide by means of a spacer. A spacer must contain at least two functional groups, one to attach to a functional group of the lipophilic substituent and the other to a functional group of the parent GLP-1 peptide.

In one embodiment, the spacer is an amino acid residue except Cys or Met, or a dipeptide such as Gly-Lys. For purposes of the present invention, the phrase "a dipeptide such as Gly-Lys" means any combination of two amino acids except Cys or Met, preferably a dipeptide wherein the C-terminal amino acid residue is Lys, His or Trp, preferably Lys, and the N-terminal amino acid residue is Ala, Arg, Asp, Asn, Gly, Glu, Gln, Ile, Leu, Val, Phe, Pro, Ser, Tyr, Thr, Lys, His and Trp. Preferably, an amino group of the parent peptide forms an amide bond with a carboxylic group of the amino acid residue or dipeptide spacer, and an amino group of the amino

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acid residue or dipeptide spacer forms an amide bond with a carboxyl group of the lipophilic substituent.

Preferred spacers are lysyl, glutamyl, asparagyl, glycyl, beta-alanyl and gamma-aminobutanoyl, each of which constitutes an individual embodiment. Most preferred spacers are glutamyl and beta-alanyl. When the spacer is Lys, Glu or Asp, the carboxyl group thereof may form an amide bond with an amino group of the amino acid residue, and the amino group thereof may form an amide bond with a carboxyl group of the lipophilic substituent. When Lys is used as the spacer, a further spacer may in some instances be inserted between the ϵ -amino group of Lys and the lipophilic substituent. In one embodiment, such a further spacer is succinic acid which forms an amide bond with the ϵ -amino group of Lys and with an amino group present in the lipophilic substituent. In another embodiment such a further spacer is Glu or Asp which forms an amide bond with the ϵ -amino group of Lys and another amide bond with a carboxyl group present in the lipophilic substituent, that is, the lipophilic substituent is a N^ϵ -acylated lysine residue.

In another embodiment, the spacer is an unbranched alkane α,ω -dicarboxylic acid group having from 1 to 7 methylene groups, which spacer forms a bridge between an amino group of the parent peptide and an amino group of the lipophilic substituent. Preferably, the spacer is succinic acid.

In a further embodiment, the lipophilic substituent with the attached spacer is a group of the formula $\text{CH}_3(\text{CH}_2)_p\text{NH}-\text{CO}(\text{CH}_2)_q\text{CO}-$, wherein p is an integer from 8 to 33, preferably from 12 to 28 and q is an integer from 1 to 6, preferably 2.

In a further embodiment, the lipophilic substituent with the attached spacer is a group of the formula $\text{CH}_3(\text{CH}_2)_r\text{CO}-\text{NHCH}(\text{COOH})(\text{CH}_2)_2\text{CO}-$, wherein r is an integer from 4 to 24, preferably from 10 to 24.

In a further embodiment, the lipophilic substituent with the attached spacer is a group of the formula $\text{CH}_3(\text{CH}_2)_s\text{CO}-\text{NHCH}((\text{CH}_2)_2\text{COOH})\text{CO}-$, wherein s is an integer from 4 to 24, preferably from 10 to 24.

In a further embodiment, the lipophilic substituent is a group of the formula $\text{COOH}(\text{CH}_2)_t\text{CO}-$ wherein t is an integer from 6 to 24.

In a further embodiment, the lipophilic substituent with the attached spacer is a group of the formula $-\text{NHCH}(\text{COOH})(\text{CH}_2)_4\text{NH}-\text{CO}(\text{CH}_2)_u\text{CH}_3$, wherein u is an integer from 8 to 18.

In a further embodiment, the lipophilic substituent with the attached spacer is a group of the formula $\text{CH}_3(\text{CH}_2)_v\text{CO}-\text{NH}-(\text{CH}_2)_z-\text{CO}$, wherein v is an integer from 4 to 24 and z is an integer from 1 to 6.

In a further embodiment, the lipophilic substituent with the attached spacer is a group of the formula $-\text{NHCH}(\text{COOH})(\text{CH}_2)_4\text{NH}-\text{COCH}((\text{CH}_2)_2\text{COOH})\text{NH}-\text{CO}(\text{CH}_2)_w\text{CH}_3$, wherein w is an integer from 10 to 16.

In a further embodiment, the lipophilic substituent with the attached spacer is a group of the formula $-\text{NHCH}(\text{COOH})(\text{CH}_2)_4\text{NH}-\text{CO}(\text{CH}_2)_2\text{CH}(\text{COOH})\text{NHCO}(\text{CH}_2)_x\text{CH}_3$, wherein x is zero or an integer from 1 to 22, preferably 10 to 16.

In yet another embodiment the GLP-1 agonist is Arg^{34} , $\text{Lys}^{26}(\text{N}^\epsilon-(\gamma\text{-Glu}(\text{N}^\alpha\text{-hexadecanoyl})))\text{-GLP-1(7-37)}$.

In yet another embodiment the GLP-1 agonist is selected from the group consisting of $\text{Gly}^8\text{-GLP-1(7-36)-amide}$, $\text{Gly}^8\text{-GLP-1(7-37)}$, $\text{Val}^8\text{-GLP-1(7-36)-amide}$, $\text{Val}^8\text{-GLP-1(7-37)}$, $\text{Val}^8\text{Asp}^{22}\text{-GLP-1(7-36)-amide}$, $\text{Val}^8\text{Asp}^{22}\text{-GLP-1(7-37)}$, $\text{Val}^8\text{Glu}^{22}\text{-GLP-1(7-36)-amide}$, $\text{Val}^8\text{Glu}^{22}\text{-GLP-1(7-37)}$, $\text{Val}^8\text{Lys}^{22}\text{-GLP-1(7-36)-amide}$, $\text{Val}^8\text{Lys}^{22}\text{-GLP-1(7-37)}$, $\text{Val}^8\text{Arg}^{22}\text{-GLP-1(7-36)-amide}$, $\text{Val}^8\text{Arg}^{22}\text{-GLP-1(7-37)}$,

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Val⁸His²²-GLP-1(7-36)-amide, Val⁸His²²-GLP-1(7-37), analogues thereof and derivatives of any of these.

In yet another embodiment the GLP-1 agonist is selected from the group consisting of Arg²⁶-GLP-1(7-37); Arg³⁴-GLP-1(7-37); Lys³⁶-GLP-1(7-37); Arg^{26,34}Lys³⁶-GLP-1(7-37); Arg^{26,34}-GLP-1(7-37); Arg^{26,34}Lys⁴⁰-GLP-1(7-37); Arg²⁶Lys³⁶-GLP-1(7-37); Arg³⁴Lys³⁶-GLP-1(7-37); Val⁸Arg²²-GLP-1(7-37); Met⁸Arg²²-GLP-1(7-37); Gly⁸His²²-GLP-1(7-37); Val⁸His²²-GLP-1(7-37); Met⁸His²²-GLP-1(7-37); His³⁷-GLP-1(7-37); Gly⁸-GLP-1(7-37); Val⁸-GLP-1(7-37); Met⁸-GLP-1(7-37); Gly⁸Asp²²-GLP-1(7-37); Val⁸Asp²²-GLP-1(7-37); Met⁸Asp²²-GLP-1(7-37); Gly⁸Glu²²-GLP-1(7-37); Val⁸Glu²²-GLP-1(7-37); Met⁸Glu²²-GLP-1(7-37); Gly⁸Lys²²-GLP-1(7-37); Val⁸Lys²²-GLP-1(7-37); Met⁸Lys²²-GLP-1(7-37); Gly⁸Arg²²-GLP-1(7-37); Val⁸Lys²²His³⁷-GLP-1(7-37); Gly⁸Glu²²His³⁷-GLP-1(7-37); Val⁸Glu²²His³⁷-GLP-1(7-37); Met⁸Glu²²His³⁷-GLP-1(7-37); Gly⁸Lys²²His³⁷-GLP-1(7-37); Met⁸Lys²²His³⁷-GLP-1(7-37); Gly⁸Arg²²His³⁷-GLP-1(7-37); Val⁸Arg²²His³⁷-GLP-1(7-37); Met⁸Arg²²His³⁷-GLP-1(7-37); Gly⁸His²²His³⁷-GLP-1(7-37); Val⁸His²²His³⁷-GLP-1(7-37); Gly⁸His³⁷-GLP-1(7-37); Val⁸His³⁷-GLP-1(7-37); Met⁸His³⁷-GLP-1(7-37); Gly⁸Asp²²His³⁷-GLP-1(7-37); Val⁸Asp²²His³⁷-GLP-1(7-37); Met⁸Asp²²His³⁷-GLP-1(7-37); Arg²⁶-GLP-1(7-36)-amide; Arg³⁴-GLP-1(7-36)-amide; Lys³⁶-GLP-1(7-36)-amide; Arg^{26,34}Lys³⁶-GLP-1(7-36)-amide; Arg^{26,34}-GLP-1(7-36)-amide; Arg^{26,34}Lys⁴⁰-GLP-1(7-36)-amide; Arg²⁶Lys³⁶-GLP-1(7-36)-amide; Arg³⁴Lys³⁶-GLP-1(7-36)-amide; Gly⁸-GLP-1(7-36)-amide; Val⁸-GLP-1(7-36)-amide; Met⁸-GLP-1(7-36)-amide; Gly⁸Asp²²-GLP-1(7-36)-amide; Gly⁸Glu²²His³⁷-GLP-1(7-36)-amide; Val⁸Asp²²-GLP-1(7-36)-amide; Met⁸Asp²²-GLP-1(7-36)-amide; Gly⁸Glu²²-GLP-1(7-36)-amide; Val⁸Glu²²-GLP-1(7-36)-amide; Met⁸Glu²²-GLP-1(7-36)-amide; Gly⁸Lys²²-GLP-1(7-36)-amide; Val⁸Lys²²-GLP-1(7-36)-amide; Met⁸Lys²²-GLP-1(7-36)-amide; Gly⁸His²²His³⁷-GLP-1(7-36)-amide; Gly⁸Arg²²-GLP-1(7-36)-amide; Val⁸Arg²²-GLP-1(7-36)-amide; Met⁸Arg²²-GLP-1(7-36)-amide; Gly⁸His²²-GLP-1(7-36)-amide; Val⁸His²²-GLP-1(7-36)-amide; Met⁸His²²-GLP-1(7-36)-amide; His³⁷-GLP-1(7-36)-amide; Val⁸Arg²²His³⁷-GLP-1(7-36)-amide; Met⁸Arg³⁷-GLP-1(7-36)-amide; Gly⁸His³⁷-GLP-1(7-36)-amide; Val⁸His³⁷-GLP-1(7-36)-amide; Met⁸His³⁷-GLP-1(7-36)-amide; Gly⁸Asp²²His³⁷-GLP-1(7-36)-amide; Val⁸Asp²²His³⁷-GLP-1(7-36)-amide; Val⁸Glu²²His³⁷-GLP-1(7-36)-amide; Met⁸Glu²²His³⁷-GLP-1(7-36)-amide; Gly⁸Lys²²His³⁷-GLP-1(7-36)-amide; Val⁸Lys²²His³⁷-GLP-1(7-36)-amide; Met⁸Lys²²His³⁷-GLP-1(7-36)-amide; Gly⁸Arg²²His³⁷-GLP-1(7-36)-amide; Val⁸Arg²²His³⁷-GLP-1(7-36)-amide; Met⁸Arg²²His³⁷-GLP-1(7-36)-amide; and derivatives thereof.

In yet another embodiment the GLP-1 agonist is selected from the group consisting of Val⁸Trp¹⁹Glu²²-GLP-1(7-37), Val⁸Glu²²Val²⁵-GLP-1(7-37), Val⁸Tyr¹⁶Glu²²-GLP-1(7-37), Val⁸Trp¹⁶Glu²²-GLP-1(7-37), Val⁸Leu¹⁶Glu²²-GLP-1(7-37), Val⁸Tyr¹⁸Glu²²-GLP-1(7-37), Val⁸Glu²²His³⁷-GLP-1(7-37), Val⁸Glu²²Ile³³-GLP-1(7-37), Val⁸Trp¹⁶Glu²²Val²⁵Ile³³-GLP-1(7-37), Val⁸Trp¹⁶Glu²²Ile³³-GLP-1(7-37), Val⁸Glu²²Val²⁵Ile³³-GLP-1(7-37), Val⁸Trp¹⁶Glu²²Val²⁵-GLP-1(7-37), analogues thereof and derivatives of any of these.

In yet another embodiment the GLP-1 agonist is exendin-4 or exendin-3, an exendin-4 or exendin-3 analogue or a derivative of any of these.

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Examples of exendins as well as analogues, derivatives, and fragments thereof to be included within the present invention are those disclosed in WO 97/46584, U.S. Pat. No. 5,424, 286 and WO 01/04156. U.S. Pat. No. 5,424,286 describes a method for stimulating insulin release with an exendin polypeptide. The exendin polypeptides disclosed include HGEFTFTSDLKQMEEEEAVRLFIEWLKNNGGX; wherein X=P or Y, and HX1X2GTFITSDLKQMEEEEAVRLFIEWLKNNGPSSGAPPPS; wherein X1X2=SD (exendin-3) or GE (exendin-4)). WO 97/46584 describes truncated versions of exendin peptide(s). The disclosed peptides increase secretion and biosynthesis of insulin, but reduce those of glucagon. WO 01/04156 describes exendin-4 analogues and derivatives as well as the preparation of these molecules. Exendin-4 analogues stabilized by fusion to serum albumin or Fc portion of an Ig are disclosed in WO 02/46227.

In one embodiment, the exendin-4 analogue is HGEFTFTSDLKQMEEEEAVRLFIEWLKNNGPSS-GAPPSKKKKKKK-amide.

Where the peptide to be included in the formulation of the invention is a GLP-1 agonist, the GLP-1 agonist is present in a concentration from about 0.1 mg/ml to about 100 mg/ml, more preferably in a concentration from about 0.1 mg/ml to about 50 mg/ml, and most preferably in a concentration of from about 0.1 mg/ml to about 10 mg/ml.

In another embodiment, the peptide to be included in the formulation of the invention is insulin, where "insulin" is understood to mean human insulin, [where "human insulin" means insulin having the amino acid sequence shown in DSHW Nicol and L F Smith: *Nature*, (1960) 4736:483-485, which is hereby incorporated by reference], human insulin analogs, human insulin derivatives or mixtures thereof, where examples of insulin analogs and derivatives are those disclosed in EP 0 792 290 (Novo Nordisk A/S), EP 0 214 826 and EP 0 705 275 (Novo Nordisk A/S), U.S. Pat. No. 5,504,188 (Eli Lilly), EP 0 368 187 (Aventis), U.S. Pat. Nos. 5,750,497 and 6,011,007, EP 375437 and EP 383472 and where such insulins may include, but are not limited to, NPH insulin, Lys β29 (Nε-tetradecanoyl) des(B30) human insulin, Lys^{B29}-(Nε-(γ-glutamyl-N^α-lithocholyl) des(B30) human insulin, N^{L29}-octanoyl insulin, 30/70 mixtures of prompt insulin zinc (SEMILENTE®) with extended insulin zinc (ULTRALENTE®), sold commercially as LENTE®, insulin glargine (LANTUS®) or extended insulin zinc (ULTRALENTE®), Lys^{B28} Pro^{B29} human insulin (HUMALOG®), Asp^{B28} human insulin, insulin aspart (NOVOLOG®), or a 30/70 mixture of insulin aspart and insulin aspart protamine (NOVOMIX®).

In one embodiment, the insulin is a derivative of human insulin or a human insulin analogue where the derivative contains at least one lysine residue and a lipophilic substituent is attached to the epsilon amino group of the lysine residue.

In one embodiment, the lysine residue to which the lipophilic substituent is attached is present at position B28 of the insulin peptide.

In an alternative embodiment, the lysine residue to which the lipophilic substituent is attached is present at position B29 of the insulin peptide.

In yet another embodiment, lipophilic substituent is an acyl group corresponding to a carboxylic acid having at least 6 carbon atoms.

In another preferred embodiment, the lipophilic substituent is an acyl group, branched or unbranched, which corresponds to a carboxylic acid having a chain of carbon atoms 8 to 24 atoms long.

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In another preferred embodiment, the lipophilic substituent is an acyl group corresponding to a fatty acid having at least 6 carbon atoms.

In another preferred embodiment, the lipophilic substituent is an acyl group corresponding to a linear, saturated carboxylic acid having from 6 to 24 carbon atoms.

In another preferred embodiment, the lipophilic substituent is an acyl group corresponding to a linear, saturated carboxylic acid having from 8 to 12 carbon atoms.

In another preferred embodiment, the lipophilic substituent is an acyl group corresponding to a linear, saturated carboxylic acid having from 10 to 16 carbon atoms.

In another preferred embodiment, the lipophilic substituent is an oligo oxyethylene group comprising up to 10, preferably up to 5, oxyethylene units.

In another preferred embodiment, the lipophilic substituent is an oligo oxypropylene group comprising up to 10, preferably up to 5, oxypropylene units.

In one preferred embodiment, the invention relates to a human insulin derivative in which the B30 amino acid residue is deleted or is any amino acid residue which can be coded for by the genetic code except Lys, Arg and Cys; the A21 and the B3 amino acid residues are, independently, any amino acid residues which can be coded for by the genetic code except Lys, Arg and Cys; Phe^{B1} may be deleted; the □-amino group of Lys^{B29} has a lipophilic substituent which comprises at least 6 carbon atoms; and 2-4 Zn²⁺ ions may be bound to each insulin hexamer with the proviso that when B30 is Thr or Ala and A21 and B3 are both Asn, and Phe^{B1} is not deleted, then 2-4 Zn²⁺ ions are bound to each hexamer of the insulin derivative.

In another preferred embodiment, the invention relates to a human insulin derivative in which the B30 amino acid residue is deleted or is any amino acid residue which can be coded for by the genetic code except Lys, Arg and Cys; the A21 and the B3 amino acid residues are, independently, any amino acid residues which can be coded for by the genetic code except Lys, Arg and Cys, with the proviso that if the B30 amino acid residue is Ala or Thr, then at least one of the residues A21 and B3 is different from Asn; Phe^{B1} may be deleted; and the □-amino group of Lys^{B29} has a lipophilic substituent which comprises at least 6 carbon atoms.

In another preferred embodiment, the invention relates to a human insulin derivative in which the B30 amino acid residue is deleted or is any amino acid residue which can be coded for by the genetic code except Lys, Arg and Cys; the A21 and the B3 amino acid residues are, independently, any amino acid residues which can be coded for by the genetic code except Lys, Arg and Cys; Phe^{B1} may be deleted; the □-amino group of Lys^{B29} has a lipophilic substituent which comprises at least 6 carbon atoms; and 2-4 Zn²⁺ ions are bound to each insulin hexamer.

Where the peptide to be included in the formulation of the invention is an insulin, the insulin is present in a concentration from about 0.5 mg/ml to about 20 mg/ml, more preferably in a concentration from about 1 mg/ml to about 15 mg/ml.

In another embodiment, the peptide to be included in the formulations of the invention is hGH or Met-hGH.

Where the peptide to be included in the formulation of the invention is hGH or Met-hGH, the hGH or Met-hGH is present in a concentration from about 0.5 mg/ml to about 50 mg/ml, more preferably in a concentration from about 1 mg/ml to about 10 mg/ml.

In yet another embodiment, the peptide to be included in the formulations of the invention is GLP-2 or an analogue or derivative thereof.

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Where the peptide to be included in the formulation of the invention is GLP-2 or an analogue or derivative thereof, the GLP-2 or an analogue or derivative thereof is present in a concentration from about 1 mg/ml to about 100 mg/ml, more preferably in a concentration from about 1 mg/ml to about 10 mg/ml.

In yet a further embodiment, the peptide to be included in the formulations of the invention is Factor VII or Factor VIIa or an analogue or derivative thereof.

Where the peptide to be included in the formulation of the invention is Factor VII or Factor VIIa or an analogue or derivative thereof, the Factor VII or Factor VIIa or an analogue or derivative thereof is present in a concentration from about 0.1 mg/ml to about 10 mg/ml, more preferably in a concentration from about 0.5 mg/ml to about 5 mg/ml.

In one embodiment, the final concentration of propylene glycol in the formulations of the invention is from about 1 to about 50 mg/ml.

In another embodiment, the final concentration of propylene glycol in the formulations of the invention is from about 5 to about 25 mg/ml.

In yet another embodiment, the final concentration of propylene glycol in the formulations of the invention is from about 8 to about 16 mg/ml.

In yet a further embodiment, the final concentration of propylene glycol in the formulations of the invention is from about 13 to about 15 mg/ml.

In still another embodiment, the final concentration of propylene glycol in the formulations of the invention is from about 13.5 to about 14.5 mg/ml.

In another embodiment of the invention, the formulation has a pH in the range from about 7.0 to about 9.5 where the term "about" as used in connection with pH means + or -0.1 pH units from the stated number.

In a further embodiment of the invention, the formulation has a pH in the range from about 7.0 to about 8.0.

In yet a further embodiment of the invention, the formulation has a pH in the range from about 7.2 to about 8.0.

In a further embodiment of the invention, the formulation has a pH in the range from about 7.0 to about 8.3.

In yet a further embodiment of the invention, the formulation has a pH in the range from about 7.3 to about 8.3.

In a preferred embodiment of the invention, the formulations contain, in addition to a peptide and propylene glycol, a buffer and/or a preservative.

Where a buffer is to be included in the formulations of the invention, the buffer is selected from the group consisting of sodium acetate, sodium carbonate, citrate, glycylglycine, histidine, glycine, lysine, arginin, sodium dihydrogen phosphate, disodium hydrogen phosphate, sodium phosphate, and tris(hydroxymethyl)-aminomethan, or mixtures thereof. Each one of these specific buffers constitutes an alternative embodiment of the invention. In a preferred embodiment of the invention the buffer is glycylglycine, sodium dihydrogen phosphate, disodium hydrogen phosphate, sodium phosphate or mixtures thereof.

Where a pharmaceutically acceptable preservative is to be included in the formulations of the invention, the preservative is selected from the group consisting of phenol, m-cresol, methyl p-hydroxybenzoate, propyl p-hydroxybenzoate, 2-phenoxyethanol, butyl p-hydroxybenzoate, 2-phenylethanol, benzyl alcohol, chlorobutanol, and thiomerosal, or mixtures thereof. Each one of these specific preservatives constitutes an alternative embodiment of the invention. In a preferred embodiment of the invention the preservative is phenol or m-cresol.

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In a further embodiment of the invention the preservative is present in a concentration from about 0.1 mg/ml to about 50 mg/ml, more preferably in a concentration from about 0.1 mg/ml to about 25 mg/ml, and most preferably in a concentration from about 0.1 mg/ml to about 10 mg/ml

The use of a preservative in pharmaceutical compositions is well-known to the skilled person. For convenience reference is made to Remington: *The Science and Practice of Pharmacy*, 19th edition, 1995.

In a further embodiment of the invention the formulation may further comprise a chelating agent where the chelating agent may be selected from salts of ethylenediaminetetraacetic acid (EDTA), citric acid, and aspartic acid, and mixtures thereof. Each one of these specific chelating agents constitutes an alternative embodiment of the invention.

In a further embodiment of the invention the chelating agent is present in a concentration from 0.1 mg/ml to 5 mg/ml. In a further embodiment of the invention the chelating agent is present in a concentration from 0.1 mg/ml to 2 mg/ml. In a further embodiment of the invention the chelating agent is present in a concentration from 2 mg/ml to 5 mg/ml.

The use of a chelating agent in pharmaceutical compositions is well-known to the skilled person. For convenience reference is made to Remington: *The Science and Practice of Pharmacy*, 19th edition, 1995.

In a further embodiment of the invention the formulation may further comprise a stabilizer selected from the group of high molecular weight polymers or low molecular compounds where such stabilizers include, but are not limited to, polyethylene glycol (e.g. PEG 3350), polyvinylalcohol (PVA), polyvinylpyrrolidone, carboxymethylcellulose, different salts (e.g. sodium chloride), L-glycine, L-histidine, imidazole, arginine, lysine, isoleucine, aspartic acid, tryptophan, threonine and mixtures thereof. Each one of these specific stabilizers constitutes an alternative embodiment of the invention. In a preferred embodiment of the invention the stabilizer is selected from the group consisting of L-histidine, imidazole and arginine.

In a further embodiment of the invention the high molecular weight polymer is present in a concentration from 0.1 mg/ml to 50 mg/ml. In a further embodiment of the invention the high molecular weight polymer is present in a concentration from 0.1 mg/ml to 5 mg/ml. In a further embodiment of the invention the high molecular weight polymer is present in a concentration from 5 mg/ml to 10 mg/ml. In a further embodiment of the invention the high molecular weight polymer is present in a concentration from 0 mg/ml to 20 mg/ml. In a further embodiment of the invention the high molecular weight polymer is present in a concentration from 20 mg/ml to 30 mg/ml. In a further embodiment of the invention the high molecular weight polymer is present in a concentration from 30 mg/ml to 50 mg/ml.

In a further embodiment of the invention the low molecular weight compound is present in a concentration from 0.1 mg/ml to 50 mg/ml. In a further embodiment of the invention the low molecular weight compound is present in a concentration from 0.1 mg/ml to 5 mg/ml. In a further embodiment of the invention the low molecular weight compound is present in a concentration from 5 mg/ml to 10 mg/ml. In a further embodiment of the invention the low molecular weight compound is present in a concentration from 10 mg/ml to 20 mg/ml. In a further embodiment of the invention the low molecular weight compound is present in a concentration from 20 mg/ml to 30 mg/ml. In a further embodiment of the invention the low molecular weight compound is present in a concentration from 30 mg/ml to 50 mg/ml.

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The use of a stabilizer in pharmaceutical compositions is well-known to the skilled person. For convenience reference is made to Remington: *The Science and Practice of Pharmacy*, 19th edition, 1995.

In a further embodiment of the invention the formulation of the invention may further comprise a surfactant where a surfactant may be selected from a detergent, ethoxylated castor oil, polyglycolized glycerides, acetylated monoglycerides, sorbitan fatty acid esters, poloxamers, such as 188 and 407, polyoxyethylene sorbitan fatty acid esters, polyoxyethylene derivatives such as alkylated and alkoxyated derivatives (tweens, e.g. Tween-20, or Tween-80), monoglycerides or ethoxylated derivatives thereof, diglycerides or polyoxyethylene derivatives thereof, glycerol, cholic acid or derivatives thereof, lecithins, alcohols and phospholipids, glycerophospholipids (lecithins, cephalins, phosphatidyl serine), glyceroglycolipids (galactopyransoide), sphingophospholipids (sphingomyelin), and sphingoglycolipids (ceramides, gangliosides), DSS (docusate sodium, docusate calcium, docusate potassium, SDS (sodium dodecyl sulfate or sodium lauryl sulfate), dipalmitoyl phosphatidic acid, sodium caprylate, bile acids and salts thereof and glycine or taurine conjugates, ursodeoxycholic acid, sodium cholate, sodium deoxycholate, sodium taurocholate, sodium glycocholate, N-Hexadecyl-N,N-dimethyl-3-ammonio-1-propane-sulfonate, anionic (alkyl-aryl-sulphonates) monovalent surfactants, palmitoyl lysophosphatidyl-L-serine, lysophospholipids (e.g. 1-acyl-sn-glycero-3-phosphate esters of ethanolamine, choline, serine or threonine), alkyl, alkoxy (alkyl ester), alkoxy (alkyl ether)-derivatives of lysophosphatidyl and phosphatidylcholines, e.g. lauroyl and myristoyl derivatives of lysophosphatidylcholine, dipalmitoylphosphatidylcholine, and modifications of the polar head group, that is cholines, ethanolamines, phosphatidic acid, serines, threonines, glycerol, inositol, and the positively charged DODAC, DOTMA, DCP, BISHOP, lysophosphatidylserine and lysophosphatidylthreonine, zwitterionic surfactants (e.g. N-alkyl-N,N-dimethylammonio-1-propanesulfonates, 3-cholamido-1-propyldimethylammonio-1-propane-sulfonate, dodecylphosphocholine, myristoyl lysophosphatidylcholine, hen egg lysolecithin), cationic surfactants (quaternary ammonium bases) (e.g. cetyl-trimethylammonium bromide, cetylpyridinium chloride), non-ionic surfactants, polyethyleneoxide/polypropyleneoxide block copolymers (Pluronic/Tetronics, Triton X-100, Dodecyl β -D-glucopyranoside) or polymeric surfactants (Tween-40, Tween-80, Brij-35), fusidic acid derivatives—(e.g. sodium tauro-dihydrofusidate etc.), long-chain fatty acids and salts thereof C6-C12 (e.g. oleic acid and caprylic acid), acylcarnitines and derivatives, N $^{\alpha}$ -acylated derivatives of lysine, arginine or histidine, or side-chain acylated derivatives of lysine or arginine, N $^{\alpha}$ -acylated derivatives of dipeptides comprising any combination of lysine, arginine or histidine and a neutral or acidic amino acid, N $^{\alpha}$ -acylated derivative of a tripeptide comprising any combination of a neutral amino acid and two charged amino acids, or the surfactant may be selected from the group of imidazoline derivatives, or mixtures thereof. Each one of these specific surfactants constitutes an alternative embodiment of the invention.

The use of a surfactant in pharmaceutical compositions is well-known to the skilled person. For convenience reference is made to Remington: *The Science and Practice of Pharmacy*, 19th edition, 1995.

The formulations of the invention may be prepared by conventional techniques, e.g. as described in Remington's *Pharmaceutical Sciences*, 1985 or in Remington: *The Science and Practice of Pharmacy*, 19th edition, 1995, where

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such conventional techniques of the pharmaceutical industry involve dissolving and mixing the ingredients as appropriate to give the desired end product.

As mentioned above, in a preferred embodiment, the formulations of the invention contain, in addition to a peptide and propylene glycol, a buffer and/or a preservative.

In one embodiment, the method for preparing such a peptide formulation comprises:

- a) preparing a first solution by dissolving preservative, propylene glycol and buffer in water;
- b) preparing a second solution by dissolving the peptide in water;
- c) mixing the first and second solutions; and
- d) adjusting the pH of the mixture in c) to the desired pH.

In another embodiment, the method for preparing such a peptide formulation comprises:

- a) preparing a first solution by dissolving preservative and buffer in water;
- b) adding propylene glycol to the first solution;
- c) mixing the first solution with a second solution containing peptide dissolved in water; and
- d) adjusting the pH of the mixture in c) to the desired pH.

In yet another embodiment, the method for preparing a peptide formulation comprises:

- a) preparing a solution by dissolving preservative, buffer and propylene glycol in water;
- b) adding the peptide to the solution of step a); and
- c) adjusting the pH of the solution of step b) to the desired pH.

As the formulations of the invention are optimal for production and for use in injection devices since they exhibit reduced deposits on production equipment and reduced clogging of injection devices, the above methods of production can be used to produce peptide formulations suitable for use in production and/or for use in injection devices.

The formulations of the invention are suitable for administration to a mammal, preferably a human. The route of administration of the formulations of the invention may be any route which effectively transports the peptide contained in the formulation to the appropriate or desired site of action, such as oral, nasal, buccal, pulmonic, transdermal or parenteral.

Due to the ability of propylene glycol to reduce clogging of injection devices when compared to other isotonic agents and to mannitol in particular, in a preferred embodiment, the formulations of the invention are to be administered parenterally to a patient in need thereof. Parenteral administration may be performed by subcutaneous, intramuscular or intravenous injection by means of a syringe, optionally a pen-like syringe. Alternatively, parenteral administration can be performed by means of an infusion pump.

A further option is a composition which may be a powder or a liquid for the administration of the formulation in the form of a nasal or pulmonic spray. As a still further option, the formulation can also be administered transdermally, e.g. from a patch, optionally a iontophoretic patch, or transmucosally, e.g. buccally. The above-mentioned possible ways to administer the formulations of the invention are not to be considered as limiting the scope of the invention.

Of course, it is understood that depending on the peptide or peptides included in the formulations of the invention, the formulations may be used in methods of treatment of diseases or conditions for which use of the peptide is indicated. One skilled in the art would understand that when used in such methods of treatment, the formulations would have to be administered in amount effective to treat the condition or disease for which the peptide was being administered where

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an "effective amount" or an "amount . . . effective" is understood to mean a dosage which is sufficient in order for the treatment of the patient with the disease or condition to be treated to be effective compared to treatment without the administered dosage. It is to be understood that "an effective amount" is the effective dose to be determined by a qualified practitioner, who may titrate dosages to achieve the desired response. Factors for consideration of dose will include potency, bioavailability, desired pharmacokinetic/pharmacodynamic profiles, the condition or disease to be treated (e.g. diabetes, obesity, weight loss, gastric ulcers), patient-related factors (e.g. weight, health, age, etc.), presence of co-administered medications (e.g. insulin), time of administration, or other factors known to a medical practitioner.

The present invention also relates to a method for reducing deposits on production equipment during production of a peptide formulation, where the method comprises replacing the isotonicity agent previously utilized in said formulation with propylene glycol at a concentration of between 1-100 mg/ml.

In one embodiment, the reduction in deposits on the production equipment during production by the propylene glycol-containing formulation relative to that observed for the formulation containing the previously utilized isotonicity agent is measured by a simulated filling experiment as described in the Examples.

In another embodiment, the isotonicity agent to be replaced by propylene glycol is selected from the group consisting of sorbitol, sucrose, glycine, mannitol, lactose monohydrate, arginine, myo-inositol and dimethylsulfone.

In a further embodiment, the isotonicity agent previously utilized in said formulation is replaced with propylene glycol in a concentration of from about 1 to about 50 mg/ml.

In another embodiment, the isotonicity agent previously utilized in said formulation is replaced with propylene glycol in a concentration of from about 5 to about 25 mg/ml.

In yet another embodiment, the isotonicity agent previously utilized in said formulation is replaced with propylene glycol in a concentration of from about 8 to about 16 mg/ml.

In another embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from about 7.0 to about 9.5.

In a further embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from about 7.0 to about 8.0.

In yet a further embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from 7.2 to about 8.0.

In a further embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from about 7.0 to about 8.3.

In a further embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from 7.3 to about 8.3.

The present invention also relates to a method for reducing deposits in the final product during production of a peptide formulation, where the method comprises replacing the isotonicity agent previously utilized in said formulation with propylene glycol at a concentration of between 1-100 mg/ml.

In one embodiment, the reduction in deposits in the final product is measured by a reduction in the number of vials and/or cartridges of the propylene glycol-containing formulation that must be discarded due to deposits relative to number of vials and/or cartridges of the formulation containing the previously utilized isotonicity agent that must be discarded due to deposits.

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In another embodiment, the isotonicity agent to be replaced by propylene glycol is selected from the group consisting of sorbitol, sucrose, glycine, mannitol, lactose monohydrate, arginine, myo-inositol and dimethylsulfon.

In a further embodiment, the isotonicity agent previously utilized in said formulation is replaced with propylene glycol in a concentration of from about 1 to about 50 mg/ml.

In another embodiment, the isotonicity agent previously utilized in said formulation is replaced with propylene glycol in a concentration of from about 5 to about 25 mg/ml.

In yet another embodiment, the isotonicity agent previously utilized in said formulation is replaced with propylene glycol in a concentration of from about 8 to about 16 mg/ml.

In another embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from about 7.0 to about 9.5.

In a further embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from about 7.0 to about 8.0.

In yet a further embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from 7.2 to about 8.0.

In a further embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from about 7.0 to about 8.3.

In a further embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from 7.3 to about 8.3.

The present invention further relates to a method for reducing the clogging of injection devices by a peptide formulation, where the method comprises replacing the isotonicity agent previously utilized in said formulation with propylene glycol at a concentration of between 1-100 mg/ml.

In one embodiment, the reduction in clogging of the injection device by the propylene glycol-containing formulation relative to that observed for the formulation containing the previously utilized isotonicity agent is measured in a simulated in use study as described in the Examples.

In another embodiment, the isotonicity agent to be replaced by propylene glycol is selected from the group consisting of inositol, maltose, glycine, lactose and mannitol.

In a further embodiment, the isotonicity agent previously utilized in said formulation is replaced with propylene glycol in a concentration of from about 1 to about 50 mg/ml.

In another embodiment, the isotonicity agent previously utilized in said formulation is replaced with propylene glycol in a concentration of from about 5 to about 25 mg/ml.

In yet another embodiment, the isotonicity agent previously utilized in said formulation is replaced with propylene glycol in a concentration of from about 8 to about 16 mg/ml.

In another embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from about 7.0 to about 9.5.

In a further embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from about 7.0 to about 8.0.

In yet a further embodiment of the invention, the propylene glycol-containing formulation has a pH in the range from 7.2 to about 8.0.

All scientific publications and patents cited herein are specifically incorporated by reference. The following examples illustrate various aspects of the invention but are in no way intended to limit the scope thereof.

EXAMPLES

Example 1

As laboratory experiments have shown that with regards to clogging of needles and deposits on needles, formulations

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without peptide ("placebo") give the same conclusions as formulations with peptide at 0.3-5.0 mg/ml, the screening studies in Example 1 have been done using placebo except where indicated otherwise.

Preparation of Formulations with Different Isotonic Agents

Preservative (5.5 mg/ml phenol) and buffer 1.24 mg/ml disodium hydrogen phosphate, dihydrate) were dissolved in water and the isotonic agent was added while stirring. pH was adjusted to pH 7.9 using Sodium Hydroxide and/or Hydrochloric acid. Finally, the formulation was filtered through a 0.22 μ m filter. The isotonic agents tested in each formulation and their concentrations are shown in Table 1.

TABLE 1

Composition of the tested formulations

Formulation no.	Tonicity modifier
1	Glucose monohydrate (38.0 mg/ml)
2	Laktose monohydrate (65.0 mg/ml)
3	Maltose (67.2 mg/ml)
4	Glycine (15.1 mg/ml)
5	Polyethylenglycol 400 (77.5 mg/ml)
6	L-arginin (24.6 mg/ml)
7	Myo-Inositol (35.2 mg/ml)
8	Propylene glycol (13.7 mg/ml)
9	Dimethylsulfon (18 mg/ml)
10	Mannitol (35.9 mg/ml)
11	Sorbitol (39.5 mg/ml)
12	Xylitol (39.5 mg/ml)
13	Sucrose (79.1 mg/ml)
14	Glycerol (16 mg/ml)

Osmolarity

The osmolarity of the different placebo formulations was determined and the results are shown in Table 2.

An isotonic solution has an osmolarity of around 0.286 osmol/L. As can be seen from Table 2 three of the formulations (PEG 400, sucrose and xylitol) are more than 20% from being isotonic (0.229-0.343 osmol/l), however for these kind of experiments the osmolarity is not expected to influence the results, though, the tonicity of the formulations should be adjusted in future experiments.

TABLE 2

The measured osmolarity of the formulations

Formulation no.	Isotonic agent	Osmolarity
1	Glucose monohydrate (38.0 mg/ml)	0.315
2	Laktose monohydrate (65.0 mg/ml)	0.283
3	Maltose (67.2 mg/ml)	0.306
4	Glycine (15.1 mg/ml)	0.286
5	Polyethylenglykol 400 (77.5 mg/ml)	0.370
6	L-arginin(24.6 mg/ml)	0.318
7	Myo-Inositol (35.2 mg/ml)	0.285
8	Propylene glycol (13.7 mg/ml)	0.268
9	Dimethylsulfon (18 mg/ml)	0.274
10	Mannitol (35.9 mg/ml)	0.284
11	Sorbitol (39.5 mg/ml)	0.310
12	Xylitol (39.5 mg/ml)	0.351
13	Sucrose (79.1 mg/ml)	0.346
14	Glycerol (16 mg/ml)	0.262

Drop Test

A droplet of each formulation is placed on a microscope slide and let to dry. The deposit is visually examined by eye and light microscope.

A photograph of the dried droplets of some of the formulations is shown in FIG. 1. In this figure it is clearly observed that mannitol cause deposits on the microscope slide when let

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to dry. No deposits were observed for sorbitol, xylitol, sucrose and glycerol. The droplet on the far right (Form 1) contains mannitol and Arg³⁴, Lys²⁶(N^ε-(γ-Glu(N^α-hexadecanoyl)))-GLP-1(7-37).

In FIG. 2 the candidates causing the most deposits on the microscope slide are shown. For comparison glycerol, which does not cause deposits, is shown (mannitol, arginine, inositol).

Clogging Test

In this test 10 NOVOPENS® 1.5 ml mounted with NOVOFINE 30® G (G 30 needle) were tested for each formulation, 5 of them placed in upright and 5 in horizontal position. The Pensystems were stored at room temperature in between testing. Each day the needle was examined for deposits and an air shot was performed prior to injection into a tissue. Degree of resistance and clogging, if any, was noted. Injections were made on a daily basis with the same needle, and this was done for 9 working days for all the formulations.

The results from the clogging test are shown in Table 3.

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three categories. 1. Those isotonic agents that do not cause deposits on the filling equipment: Xylitol, glycerol, glucose monohydrate, maltose, PEG 400 and propylene glycol. 2. Those isotonic agent that cause few deposits and have superior filling properties compared to mannitol: Sorbitol, sucrose and glycine. 3. Those isotonic agent that are comparable or worse than mannitol: Mannitol, lactose monohydrate, arginine, myo-inositol and dimethylsulfon.

Conclusion

In the simulated filling experiment xylitol, glycerol, glucose, maltose, PEG 400, propylene glycol, sorbitol, sucrose and glycine were found to be suitable replacements candidates for mannitol. However, as glucose is a reducing saccharide, and therefore is able to initiate unwanted degradation in the formulation, this tonicity modifier is ruled out. Furthermore, maltose is ruled out due to clogging of needles. This leads to the following candidates: glycerol, xylitol, sorbitol, sucrose, glycine, propylene glycol and PEG 400, which are found to have suitable properties as replacements candidates

TABLE 3

Clogging test in NovoPen 1.5 using 30G NovoFine

Isotonic agent (no. of observations)	Some resistance	Resistance	Much resistance	Clogged	Drop at top of needle	Dried drop at needle top	Gel-like drop on needle	Deposits on needle
Mannitol (90)	10	0	0	0	0	2	0	43
Glycerol (90)	13	0	0	0	1	0	3	0
Sucrose (90)	23	0	0	0	0	0	21	0
Propylene glycol (90)	20	0	0	0	0	0	0	0
PEG 400 (90)	25	1	0	0	12 (5 at needle)	0	0	0
arginin (90)	26	2	0	0	3 (2 at needle)	1	0	0
Xylitol (90)	14	0	0	0	5	0	0	0
Dimethylsulfon (90)	21	0	0	0	4	0	0	0
sorbitol (90)	12	0	0	0	9	1	0	1
Myo-inositol (90)	20	1	2	6	6	0	0	47
Glucose (90)	32	11	5	0	16 (7 at needle)	1	0	(1 at needle)
glycine (90)	41	9	2	0	1 (2 at needle)	0	0	31 (2 at needle)
maltose (90)	35	8	7	4	16 (6 at needle)	0	0	1 (5 at needle)
laktose (90)	44	10	8	0	5	0	0	31 (2 at needle)

In Table 3 and in FIG. 3 it was observed that inositol and maltose clogged the needle. For comparison glycerol which does not clog the needle is shown in FIG. 3. In FIG. 4, and in Table 3, it was observed that formulations containing glycine, lactose and mannitol gave rise to a lot of deposits on the needle. For glycine, the deposits were a droplet deposited down the needle, whereas for lactose and mannitol the deposits occurred at the top of the needle.

Simulated Filling

1 L of each formulation was subjected to a simulated filling experiment which lasted for 24 hours. After 24 hours the filling equipment was inspected for the presence of deposits.

Based on the results from the simulated filling studies (data not shown), the placebo formulations can be divided into

for mannitol in peptide formulations with regards to drop test, clogging of needles and simulated filling.

However, on the basis of the following considerations, propylene glycol was chosen as the isotonic agent over the other candidates to be further investigated in head to head comparison studies with mannitol:

- propylene glycol was observed to have no influence on the physical and chemical stability of Arg³⁴, Lys²⁶(N^ε-(γ-Glu(N^α-hexadecanoyl)))-GLP-1(7-37)-containing formulations;
- propylene glycol was observed to have no influence on antimicrobial preservative testing; and
- use of propylene glycol would no require that further toxicity studies be tested

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Example 2

Comparison of Mannitol and Propylene Glycol-Containing Placebo Formulations in Simulated Filling Studies and Simulated Use Studies

Preparation of Formulations

Preservative and buffer were dissolved in water and the isotonic agent was added while stirring. pH was adjusted to the aimed pH using Sodium Hydroxide and/or Hydrochloric acid. Finally, the formulation was filtered through a 0.22 μ m filter. The compositions of the formulations were as follows:

Disodium hydrogen phosphate, dihydrate: 1.42 mg/ml

Phenol: 5.5 mg/ml

Propylene glycol or mannitol: 13.7 or 35.9 mg/ml

Water for Injection: up to 1.0 ml.

pH: 7.90

Simulated Filling Study

A simulated filling study lasting 24 hours was performed as described in Example 1 and after 24 hours, the filling equipment was inspected for the presence of deposits. No deposits were observed on the filling equipment for the propylene glycol formulation. By comparison, after 24 hours, a lot of deposits were observed on the filling equipment for the mannitol formulation (see FIG. 6).

Simulated in Use Study

For the simulated in use study, a clogging test was conducted as described in Example 1. The same needle was used during the study period of ten working days and each day, the needle was inspected for the presence of deposits. FIG. 7 shows photographs of needles dosed with the propylene glycol (top panel) or mannitol (bottom panel) containing formulations. Deposits on the needle were observed in 48% of the cases when mannitol was used as an isotonic agent whereas no deposits were observed when propylene glycol was used as the isotonic agent.

Example 3

Comparison of Propylene Glycol to Mannitol in Arg³⁴, Lys²⁶ (N^ε-(γ -Glu(N^α-hexadecanoyl)))-GLP-1(7-37) Containing Formulations

Preparation of Formulations

Preservative, isotonic agent (mannitol or propylene glycol) and buffer were dissolved in water and pH was adjusted to the desired pH. Arg³⁴, Lys²⁶(N^ε-(γ -Glu(N^α-hexadecanoyl)))-GLP-1(7-37) was dissolved in water while stirring slowly. The two solutions were then mixed and pH adjusted to the desired pH using sodium hydroxide and/or hydrochloric acid. Finally, the formulation was filtered through a 0.22 μ m filter. The compositions of the formulations were as follows:

Arg³⁴, Lys²⁶(N^ε-(γ -Glu(N^α-hexadecanoyl)))-GLP-1(7-37) (6.25 mg/ml),

Disodium hydrogen phosphate, dihydrate (1.42 mg/ml),

Phenol (5.5 mg/ml),

mannitol or propylene glycol (35.9 or 14.0 mg/ml),

Water for Injection (up to 1.0 ml),

pH: 8.15

Simulated in Use Study

For the simulated in use study, a clogging test was conducted as described in Example 1 except that a G31 needle was used. The same G31 needle was used during the study period of ten working days and each day, the needle was inspected for the presence of deposits. FIG. 7 shows photographs of needles with no deposits when dosed with the propylene glycol (bottom panel) or showing deposits when dosed with the mannitol (top panel) containing formulations.

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For the mannitol containing formulation, clogging of the needle was observed in 1 out of 10 cases on day 4, 2 out of 10 cases on day 5, 3 out of 10 cases on day 8 and 4 out of 10 cases on day 9. By comparison, no clogging of needles was observed for the propylene glycol containing formulation.

It is believed that similar results to those obtained with the above-described propylene glycol-containing formulation would also be obtained if the pH was adjusted to 7.40, 7.70 or 7.90. In addition, additional formulations which could be tested include those having the following compositions:

Buffering agents: glycylglycine (1.32 mg/ml), L-Histidine (1.55 mg/ml), Hepes (2.38 mg/ml), or bicine (1.63 mg/ml)

Preservatives: phenol (5.0 or 5.5 mg/ml), benzylalcohol (18 mg/ml) or a mixture of m-cresol and phenol (2.5/2.0 mg/ml)

Propylene glycol: 14.0 or 14.3 mg/ml

Water for injection: up to 1.0 ml

pH: 7.40, 7.70, 7.90 or 8.15

Example 4

Influence of Peptide Concentration on Clogging of Needles

Arg³⁴, Lys²⁶(N^ε-(γ -Glu(N^α-hexadecanoyl)))-GLP-1(7-37) formulations were prepared as described in Example 3 using peptide concentrations ranging from 0-5 mg/ml of Arg³⁴, Lys²⁶(N^ε-(γ -Glu(N^α-hexadecanoyl)))-GLP-1(7-37). The compositions of the formulations were as follows:

Liraglutide: 0, 0.3, 3 and 5 mg/ml

Disodium hydrogen phosphate, dihydrate: 0.71 mg/ml

Sodium dihydrogenphosphate, dihydrate: 0.62 mg/ml

Mannitol: 36.9 mg/ml

Phenol: 5.0 mg/ml

Water for injection: up to 1.0 ml

pH 7.40

A simulated in use study was conducted as in Example 3 except that a G30 needle was used and the results (data not shown) indicated that the clogging effect of the mannitol-containing formulations relative to the absence of clogging with the propylene glycol formulations was observed independent of the peptide concentration.

Example 5

Clogging of Needles in Lys β 29 (N^ε-tetradecanoyl) des(B30) Human Insulin and NovoMix 30 Formulations Containing Mannitol

Preparation Of Formulations

The Lys β 29 (N^ε-tetradecanoyl) des(B30) human insulin-containing formulation was prepared as follows:

a) Prepared a first solution by dissolving buffer, sodium chloride, preservatives (phenol and m-cresol) and mannitol in water

b) Prepared a second solution of Lys β 29 (N^ε-tetradecanoyl) des(B30) human insulin and zinc acetate dissolved in water

c) added the peptide-containing solution of step b) to the solution of step a); and

d) adjusted the pH of the solution to the desired pH

The composition of Lys β 29 (N^ε-tetradecanoyl) des(B30) human insulin-containing formulation prepared in the above manner was as follows:

Lys β 29 (N^ε-tetradecanoyl) des(B30) human insulin (2400 nmol), Phenol (1.80 mg/ml), m-cresol (2.06 mg/ml), Mannitol (30.0 mg/ml), disodiumphosphate, dihydrate (0.890 mg/ml), Sodium chloride (1.17 mg/ml), Zinc acetate (65.4 μ g/ml), water for injection (up to 1.0 ml), pH: 7.4

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The NOVOMIX® 30-containing formulation was prepared as follows:

- a) Prepared a solution by dissolving buffer, sodium chloride, phenol, mannitol and sodium hydroxide in water
- b) Prepared a solution of sodium chloride, phenol and mannitol in water
- c) Prepared a solution of protamine sulphate in water
- d) Prepared a solution of insulin, hydrochloric acid and zinc in water
- e) Solutions b), c) and d) were mixed
- f) Solution e) was added to the solution of step a)
- g) Adjusted the pH of the solution to the desired pH and crystallized at room temperature
- h) Prepared a solution by dissolving m-cresol, phenol and mannitol in water
- i) Solution h) is added to the crystalline fraction of step g); and
- j) Adjusted the pH to the desired pH

The composition of the NOVOMIX® 30-containing formulation prepared in the above manner was as follows:

Insulin aspart (100 units/ml), protamine sulphate (approx. 0.33 mg/ml), phenol (1.50 mg/ml), m-cresol (1.72 mg/ml),

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Example 6

Testing of Lys β29 (Nε-tetradecanoyl) des(B30) human insulin and NOVOMIX® 30 formulations containing propylene glycol

The preparation and composition of the Lys β29 (Nε-tetradecanoyl) des(B30) human insulin and NOVOMIX® 30 formulations will be as described in Example 5 except that mannitol will be replaced with a concentration of propylene glycol that assures tonicity. A simulated in use test will then be conducted as described in Example 5.

Based on the fact that the clogging effect of Lys β29 (Nε-tetradecanoyl) des(B30) human insulin and NOVOMIX® 30 mannitol-containing formulations was similar to that observed with Arg³⁴, Lys²⁶(Nε-(γ-Glu(Nα-hexadecanoyl)))-GLP-1(7-37) mannitol-containing formulations, it is believed that the effect of propylene glycol on the clogging effect of Lys β29 (Nε-tetradecanoyl) des(B30) human insulin and NovoMix 30-containing formulations will be similar to that observed with Arg³⁴, Lys²⁶(Nε-(γ-Glu(Nα-hexadecanoyl)))-GLP-1(7-37)-containing formulations.

SEQUENCE LISTING

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<222> LOCATION: (44)..(44)
<223> OTHER INFORMATION: Lysine at position 44 is amidated

<400> SEQUENCE: 1

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Glu Ala Val Arg Leu Phe Ile Glu Trp Leu Lys Asn Gly Gly Pro Ser
20          25          30

Ser Gly Ala Pro Pro Ser Lys Lys Lys Lys Lys Lys
35          40
```

mannitol (30.0 mg/ml), disodiumphosphate dihydrate (1.25 mg/ml), sodium chloride (0.58 mg/ml), zinc (19.6 ug/ml), water for injection (up to 1.0 ml), pH: 7.3.

Results

A simulated in use study was conducted as described in Example 3 using G31 needles where 20 needles were investigated for 10 days. The results were as follows: Clogging of needles was observed for Lys β29 (Nε-tetradecanoyl) des(B30) human insulin on day 2 (5%), day 3 (70%) and on day 4 (100%). Clogging of needles for NovoMix 30 was observed on day 3 (5%), day 4 (10%), day 5 (35%), day 6 (40%), day 8 (50%), day 9 (55%) and day 10 (80%). Thus, the effect of mannitol on the clogging of needles is independent of the type of peptide included in the formulations since a comparable clogging effect was observed with Arg³⁴, Lys²⁶(Nε-(γ-Glu(Nα-hexadecanoyl)))-GLP-1(7-37), Lys β29 (Nε-tetradecanoyl) des(B30) human insulin and NovoMix 30.

The invention claimed is:

1. A pharmaceutical formulation comprising at least one GLP-1 agonist, a disodium phosphate dihydrate buffer and propylene glycol, wherein said propylene glycol is present in said formulation in a final concentration of from about 1 mg/ml to about 100 mg/ml and wherein said formulation has a pH of from about 7.0 to about 10.0.
2. The formulation according to claim 1, wherein the concentration of propylene glycol is from about 1 mg/ml to about 50 mg/ml.
3. The formulation according to claim 1, wherein the concentration of propylene glycol is from about 5 mg/ml to about 25 mg/ml.
4. The formulation according to claim 1, wherein the concentration of propylene glycol is from about 8 mg/ml to about 16 mg/ml.
5. The formulation according to claim 1, wherein the pH of said formulation is about 7.0 to about 9.5.
6. The formulation according to claim 1, wherein the pH of said formulation is about 7.0 to about 8.3.

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7. The formulation according to claim 1, wherein the pH of said formulation is about 7.3 to about 8.3.

8. The formulation according to claim 1, further comprising a preservative.

9. The formulation according to claim 8, wherein said preservative is present in a concentration from 0.1 mg/ml to 20 mg/ml.

10. The formulation according to claim 1, wherein said GLP-1 agonist is selected from the group consisting of GLP-1(7-36)-amide, GLP-1(7-37), a GLP-1(7-36)-amide analogue, a GLP-1(7-37) analogue, or a derivative of any of these.

11. The formulation according to claim 10, wherein said GLP-1 agonist is a derivative of GLP-1(7-36) or GLP-1(7-37) or a GLP-1(7-36)-amide analogue or a GLP-1(7-37) analogue, where said derivative has a lysine residue and a lipophilic substituent attached with or without a spacer to the epsilon amino group of said lysine.

12. The formulation according to claim 11, wherein said lipophilic substituent has from 8 to 40 carbon atoms.

13. The formulation according to claim 12, wherein said spacer is an amino acid.

14. The formulation according to claim 13, wherein said GLP-1 agonist is Arg³⁴, Lys²⁶(N-ε-(γ-Glu(N-α-hexadecanoyl)))²⁶-GLP-1(7-37).

15. The formulation according to claim 1, wherein said GLP-1 agonist is selected from the group consisting of Gly⁸-GLP-1(7-36)-amide, Gly⁸-GLP-1(7-37), Val⁸-GLP-1(7-36)-amide, Val⁸-GLP-1(7-37), Val⁸Asp²²-GLP-1(7-36)-amide, Val⁸Asp²²-GLP-1(7-37), Val⁸Glu²²-GLP-1(7-36)-amide, Val⁸Glu²²-GLP-1(7-37), Val⁸Lys²²-GLP-1(7-36)-amide, Val⁸Lys²²-GLP-1(7-37), Val⁸Arg²²-GLP-1(7-36)-amide, Val⁸Arg²²-GLP-1(7-37), Val⁸His²²-GLP-1(7-36)-amide, Val⁸His²²-GLP-1(7-37), Arg³⁴GLP-1(7-37), Arg²⁶, Arg³⁴Lys³⁶GLP-1(7-36), Arg²⁶GLP-1(7-37), and Gly⁸, Arg²⁶, Arg³⁴Glu³⁷Lys³⁸GLP-1(7-38) and derivatives of any of these.

16. A method of preparing a GLP-1 agonist formulation suitable for use in an injection device, said method comprising preparing a formulation containing a GLP-1 agonist, propylene glycol, a disodium phosphate dihydrate buffer, and a preservative, wherein said propylene glycol is present in a concentration from about 1 mg/ml to about 100 mg/ml, and wherein said formulation has a pH from about 7.0 to about 10.0, and wherein said GLP-1 agonist, said propylene glycol and said buffer and preservative are mixed together to produce said formulation as follows:

- a) preparing a first solution by dissolving preservative, propylene glycol and buffer in water;
 - b) preparing a second solution by dissolving the GLP-1 agonist in water;
 - c) mixing the first and second solutions; and
- adjusting the pH of the mixture in c) to a pH of from about 7.0 to about 10.0.

17. The method according to claim 16, wherein the concentration of propylene glycol is from about 1 mg/ml to about 50 mg/ml.

18. The method according to claim 16, wherein the concentration of propylene glycol is from about 5 mg/ml to about 25 mg/ml.

19. The method according to claim 16, wherein the concentration of propylene glycol is from about 8 mg/ml to about 16 mg/ml.

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20. The method according to claim 16, wherein the pH of said formulation is about 7.0 to about 9.5.

21. The method according to claim 16, wherein the pH of said formulation is about 7.0 to about 8.0.

22. The method according to claim 16, wherein the pH of said formulation is about 7.2 to about 8.0.

23. A method for reducing deposits on production equipment during production of a GLP-1 agonist formulation, said method comprising replacing the isotonicity agent previously utilized in said formulation with propylene glycol at a concentration of between 1-100 mg/ml, and wherein said GLP-1 agonist formulation comprises a disodium phosphate dihydrate buffer.

24. The method according to claim 23, wherein the reduction in deposits on the production equipment during production by the propylene glycol-containing formulation relative to that observed for the formulation containing the previously utilized isotonicity agent is measured by a simulated filling experiment.

25. The method according to claim 23, wherein the isotonicity agent to be replaced by propylene glycol is selected from the group consisting of sorbitol, sucrose, glycine, mannitol, lactose monohydrate, arginin, myo-inositol and dimethylsulfon.

26. A method for reducing deposits in the final product during production of a GLP-1 agonist formulation, said method comprising replacing the isotonicity agent previously utilized in said formulation with propylene glycol at a concentration of between 1-100 mg/ml, and wherein said GLP-1 agonist formulation comprises a disodium phosphate dihydrate buffer.

27. The method according to claim 26, wherein the reduction in deposits in the final product is measured by a reduction in the number of vials and/or cartridges of the propylene glycol-containing formulation that must be discarded due to deposits relative to number of vials and/or cartridges of the formulation containing the previously utilized isotonicity agent that must be discarded due to deposits.

28. The method according to claim 26, wherein the isotonicity agent to be replaced by propylene glycol is selected from the group consisting of sorbitol, glycerol, sucrose, glycine, mannitol, lactose monohydrate, arginin, myo-inositol and dimethylsulfon.

29. A method for reducing the clogging of injection devices by a GLP-1 agonist formulation, said method comprising replacing the isotonicity agent previously utilized in said formulation with propylene glycol at a concentration of between 1-100 mg/ml, and wherein said GLP-1 agonist formulation comprises a disodium phosphate dihydrate buffer.

30. The method according to claim 29, wherein the reduction in clogging of the injection device by the propylene glycol-containing formulation relative to that observed for the formulation containing the previously utilized isotonicity agent is measured in a simulated in use study.

31. The method according to claim 29, wherein the isotonicity agent to be replaced by propylene glycol is selected from the group consisting of inositol, maltose, glycine, lactose and mannitol.

* * * * *

EXHIBIT B



US009265893B2

(12) **United States Patent**
Hansen et al.

(10) **Patent No.:** **US 9,265,893 B2**
(45) **Date of Patent:** **Feb. 23, 2016**

(54) **INJECTION BUTTON**

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A61M 5/315 (2006.01)

(52) **U.S. Cl.**
CPC **A61M 5/31585** (2013.01); **A61M 5/31511** (2013.01); **A61M 5/3158** (2013.01)

(58) **Field of Classification Search**

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USPC 604/181, 187, 218–231
See application file for complete search history.

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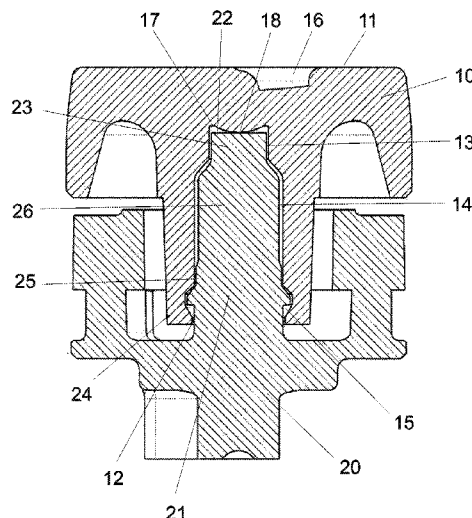
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(57) **ABSTRACT**

A push button connection for an injection device comprising a push button (10) and a driving part (20). The two parts of the push button connection are mounted to each other and is relatively rotatable to each other. In order to minimize the friction occurring between the push button and the driving part when relatively rotated forces are transmitted via a pivot bearing (18, 22). In order also to minimize the effect of forces appearing dislocated from the center line a number of radial bearings (13, 23; 14, 25) having a little friction diameter is provided.

6 Claims, 2 Drawing Sheets



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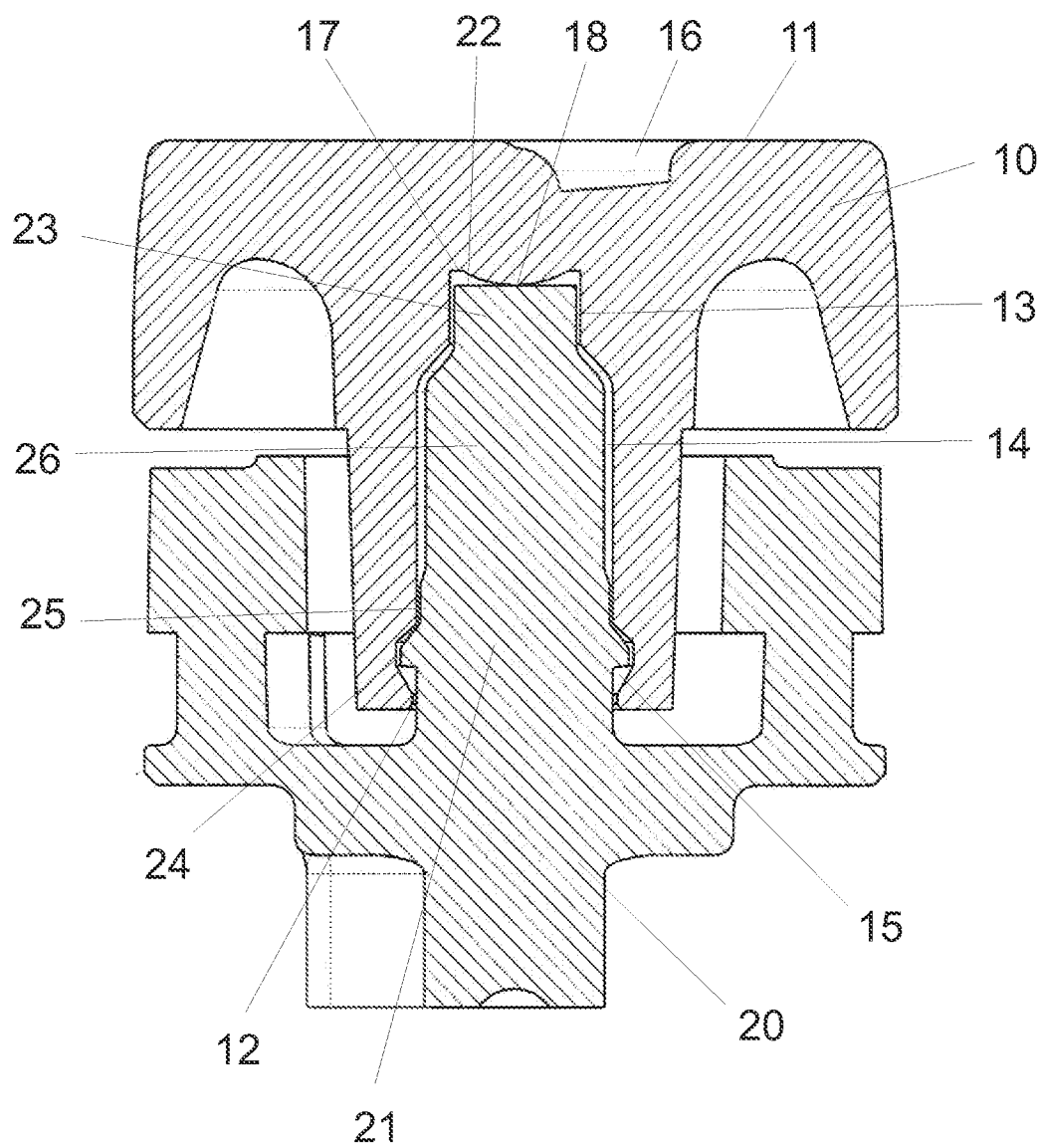


Fig. 1

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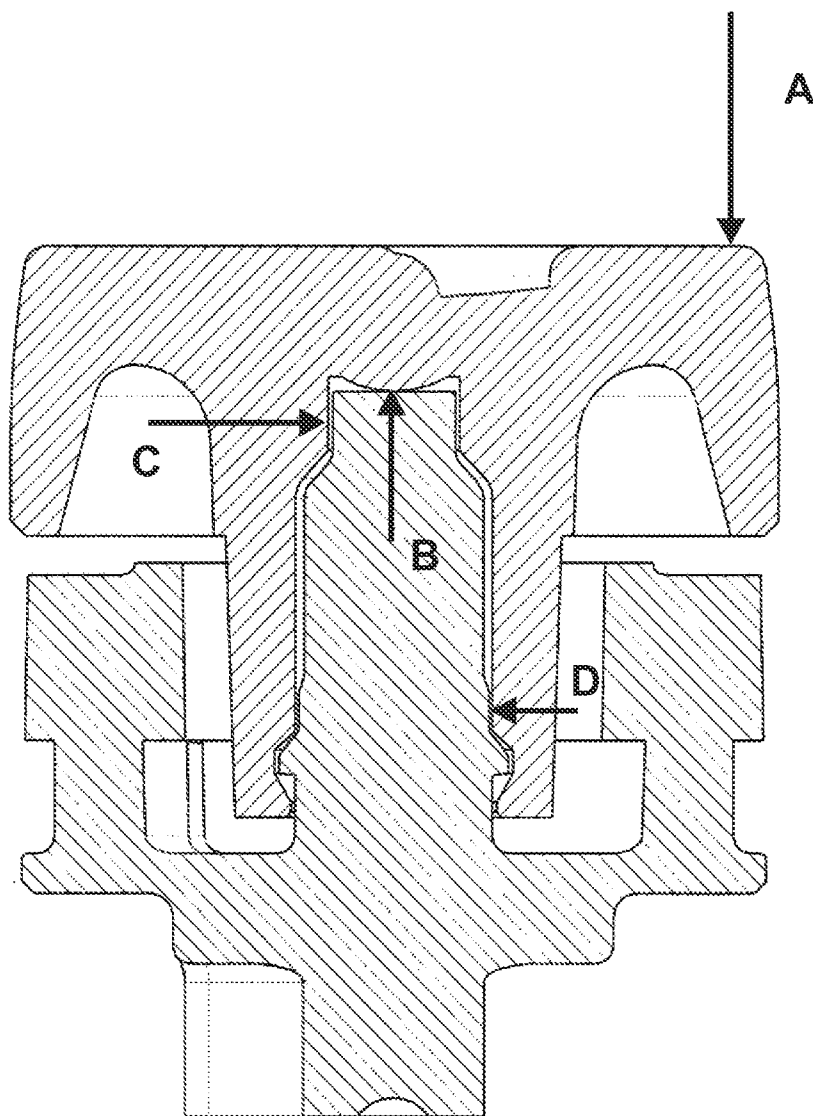


Fig. 2

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INJECTION BUTTON

CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a 35 U.S.C. §371 national stage application of International Patent Application PCT/EP2008/050624 (published as WO2008/095762), filed Jan. 21, 2008, which claimed priority of European Patent Application 07101729.7, filed Feb. 5, 2007; this application further claims priority under 35 U.S.C. §119 of U.S. Provisional Application 60/899,977, filed Feb. 7, 2007.

THE TECHNICAL FIELD OF THE INVENTION

The invention relates to a push button connection for an injection device and especially to such connection where a push button is rotated relatively to a driving member to which it is connected.

DESCRIPTION OF RELATED ART

U.S. Pat. No. 6,235,004 discloses an injection device in which according to FIG. 15-16 a dose is set by rotating the scale drum out of the housing in a threaded connection. When injecting the set dose the user pushes on the push button which forces the scale drum and the bushing to rotate together back into the housing. During this rotation of the bushing to which the push button is attached, the push button and the bushing rotates relatively to each other. The friction occurring between these relatively rotatable parts contributes to the force a user needs to apply in order to push back the bushing and the scale drum in order to inject the set dose.

U.S. Pat. No. 7,427,275 discloses an injection device in which the push button is formed with a bore encompassing a stem on a sleeve member. The push button and the stem are welded together such that the push button and the sleeve member are axially and rotatably fixed to each other.

DESCRIPTION OF THE INVENTION

It is an object of the present invention to provide a dose button connection for an injection device which minimizes the forces a user must apply to inject a dose.

When a user pushes on the injection button, the force applied is directed to the forward movement of the driving part, however, since the push button and the driving part rotate relatively to each other a friction between these rotating parts will occur. The user therefore also has to apply a force large enough to overcome this friction. One way of minimizing the force a user must apply in order to perform an injection is therefore to minimize this friction. By forming a pivot bearing between the two parts, the surface area of interaction between the two objects can be minimized and the radius of the resulting friction force can be kept at a minimum.

In order to secure the fit between the push button and the driving part and on the same time direct forces applied on the periphery of the push button to the driving part at least one radial bearing between the push button and the protrusion is formed.

Preferably one radial bearing is formed in the upper area and one is formed in the lower area both having the least possible radius of friction. In this way forces applied at in the periphery area of the push button and causing tilting of the push button on the protrusion of the driving part is properly transferred.

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If a user applies a force eccentric to the centre axis of the push button i.e. on a peripheral area of the button, the push button will tilt and a reaction torque will occur at the radial bearings. In order to minimize this force pair, which in this load case is located at the distance from the radial bearing surface to the centre axis of the system, this distance, which again equals the radius of the protrusion, must be as little as possible and the distance between the bearings as long as possible. However, in order not to make the protrusion too narrow and fragile it is preferred to balance the radius of the bearings, such that the upper bearing has the smallest diameter and the lower bearing at the root of the column shaped protrusion has a diameter large enough to resist the bending force as a result of the offset applied push button forces.

In order to assemble the push button in an irreversible manner making it difficult to disassemble, it is preferred to secure the push button at the intended position by adding a track into which a rim on the harder part is forced during the manufacture of the injection device. The necessary compliance of the push button for the assembly snap-on can be secured by selection of a soft material and/or a vertical slit in the hollow section of the geometry.

Further the materials used for the push button and the protrusion on the driving part could be materials having low internal friction, or the materials could be surface treated in order to lower the internal friction.

The push button used in the connection has a central bore dedicated to engage the protrusion provided on the driving part. The bottom of the bore is preferable formed with a pivot. This pivot bears on a surface of the protrusion thus forming a pivot bearing.

DEFINITIONS

An "injection pen" is typically an injection apparatus having an oblong or elongated shape somewhat like a pen for writing. Although such pens usually have a tubular cross-section, they could easily have a different cross-section such as triangular, rectangular or square or any variation around these geometries.

As used herein, the term "drug" is meant to encompass any drug-containing flowable medicine capable of being passed through a delivery means such as a hollow needle in a controlled manner, such as a liquid, solution, gel or fine suspension. Representative drugs includes pharmaceuticals such as peptides, proteins (e.g. insulin, insulin analogues and C-peptide), and hormones, biologically derived or active agents, hormonal and gene based agents, nutritional formulas and other substances in both solid (dispensed) or liquid form.

All references, including publications, patent applications, and patents, cited herein are incorporated by reference in their entirety and to the same extent as if each reference were individually and specifically indicated to be incorporated by reference and were set forth in its entirety herein.

All headings and sub-headings are used herein for convenience only and should not be constructed as limiting the invention in any way.

The use of any and all examples, or exemplary language (e.g. such as) provided herein, is intended merely to better illuminate the invention and does not pose a limitation on the scope of the invention unless otherwise claimed. No language in the specification should be construed as indicating any non-claimed element as essential to the practice of the invention. The citation and incorporation of patent documents herein is done for convenience only and does not reflect any view of the validity, patentability, and/or enforceability of such patent documents.

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This invention includes all modifications and equivalents of the subject matter recited in the claims appended hereto as permitted by applicable law.

BRIEF DESCRIPTION OF THE DRAWINGS

The invention will be explained more fully below in connection with a preferred embodiment and with reference to the drawings in which:

FIG. 1 Show a cross section view of the connection between a push button and a driving part.

FIG. 2 Show a cross section view of the connection and the forces occurring.

The figures are schematic and simplified for clarity, and they just show details, which are essential to the understanding of the invention, while other details are left out. Throughout, the same reference numerals are used for identical or corresponding parts.

DETAILED DESCRIPTION OF EMBODIMENT

When in the following terms as “upper” and “lower”, “right” and “left”, “horizontal” and “vertical”, “clockwise” and “counter clockwise” or similar relative expressions are used, these only refer to the appended figures and not to an actual situation of use. The shown figures are schematic representations for which reason the configuration of the different structures as well as there relative dimensions are intended to serve illustrative purposes only.

In that context it may be convenient to define that the term “distal end” in the appended figures is meant to refer to the end of the injection device carrying the injection needle whereas the term “proximal end” is meant to refer to the opposite end pointing away from the injection needle.

FIG. 1 discloses the connection between the push button 10 and the driving part 20.

When a user wants to inject a dose, which he or she has first selected, the user pushes the push button 10 which then moves the driving part 20 axially forward in the injection device. During this forward movement of the driving part 20 it also rotates usually because it is interfaced with a dose dial drum which is threadedly connected to a housing. Such injection device is described in details in EP 1.003.581. The combined axial and rotatable movement of the driving part 20 drives the set dose out from the injection device.

As the users finger pushes on the push surface 11 of the push button 10 it is unable to rotate due to the friction between the users finger and the push button 10 whereas the driving part 20 is forced to rotate due to its interface, therefore a relative rotation between the push button 10 and the driving part 20 takes place.

The push button 10 which could be manufactured from a suitable polymeric material being softer than the material from which the driving part 20 is manufactured comprises at the proximal end a push surface 11 which is contacted by the user's finger when a dose is to be injected and an opposite located cylindrical bore 12 with a circular cross section. The most proximal part 13 of the bore 12 has a smaller diameter than the remaining part 14 of the bore 12. At the distal end of the bore 12, a radial extending track 15 is provided.

The push surface 11 could be provided with a tactile cut-out 16 informing visual impaired users on the content of the injection device and the most proximal bottom surface 17 of the bore 12 is formed with a raised pointer forming a pivot 18.

The driving part 20 is provided with a protrusion 21 having a circular cross section and a top surface 22. This protrusion 21 has at its proximal end a top part 23 with a decreased

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diameter compared to the remaining part 26 of the protrusion 21. Further the protrusion 21 is provided with a radial extending rim 24 at its distal end. In the area around this rim 24, the protrusion 21 is provided with a belt 25 with a slightly raised diameter.

When the push button 10 is mounted on the protrusion 21 of the driving element 20 it is simply clicked on such that the extending rim 24 enters the track 15. This forms a connection almost impossible to disconnect since the polymeric material of the push button 10 is softer than the material from which the protrusion 21 is produced. In this position the pivot 18 formed in the most proximal bottom surface 17 of the bore 12 bears on the top surface 22 of the protrusion 21 thus forming a pivot bearing 22, 18. Further the push button 10 is radially supported by the protrusion 21 at the top part 23 forming a radial top bearing 23, 13. The belt 25 on the protrusion 21 bears on an area of the remaining part 14 of the bore 12 thus forming a radial bottom bearing 14, 25.

In FIG. 2 the push button 10 connection is disclosed with the various forces occurring when a user applies an injection force in the peripheral area of the push button 10.

When the user applies an injection force A at the peripheral area of the push button 10 a vertical reaction force B will appear at the pivot point 22, 18, at the same time a radial force C will occur at the upper radial bearing 13, 23. Since the upper radial bearing 13, 23 are located at the top part 23 having the smaller diameter, the resulting torque is relatively small. Further, a radial force D will occur at the lower radial bearing 14, 25, however due to the distance between the upper radial bearing 13, 23 and the lower radial bearing 14, 25, the force resulting on the lower radial bearing 14, 25 is relatively small.

Some preferred embodiments have been shown in the foregoing, but it should be stressed that the invention is not limited to these, but may be embodied in other ways within the subject matter defined in the following claims.

The invention claimed is:

1. A push button connection for an injection device comprising:

a push button mountable on a driving part being rotatable relatively to the push button and which push button further comprises a bore with a bottom surface and which bore surrounds a protrusion on the driving part which protrusion has a top surface and wherein a pivot bearing is formed between the bottom surface and the top surface, wherein when a user presses on the push button the force is directed toward the driving part and wherein the driving part rotates relative to the push button.

2. A push button connection according to claim 1, in which at least one radial bearing between the push button and the driving part is provided.

3. A push button connection according to claim 2, in which an upper radial bearing is provided at a top part of the protrusion and a lower radial bearing is provided at the bottom of the protrusion.

4. A push button connection according to claim 3, in which the top part of the protrusion accommodating the upper radial bearing has a diameter smaller than the diameter of the remaining part of the protrusion.

5. A push button connection according to claim 1, in which the push button is manufactured from a polymeric material being softer than the material from which the driving part is manufactured.

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6. A push button connection according to claim 1, in which the protrusion is provided with an extending rim mating with a track provided in the push button.

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